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(54) A Method of identifying unknown adeno-associated virus (AVV) sequences and a kit for the method

Verfahren zur Identifizierung von Adeno-assoziiertem Virus (AAV) Sequenzen sowie Kit zur Ausführung der Methode

Une méthode d'identification de séquences de virus adéno-associés et kit permettant d'appliquer la méthode

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#### Description

#### BACKGROUND OF THE INVENTION

[0001] Adeno-associated virus (AAV), a member of the Parvovirus family, is a small nonenveloped, icosahedral virus with single-stranded linear DNA genomes of 4.7 kilobases (kb) to 6 kb. AAV is assigned to the genus, Dependovirus, because the virus was discovered as a contaminant in purified adenovirus stocks. AAV's life cycle includes a latent phase at which AAV genomes, after infection, are site specifically integrated into host chromosomes and an infectious phase in which, following either adenovirus or herpes simplex virus infection, the integrated genomes are subsequently rescued, replicated, and packaged into infectious viruses. The properties of non-pathogenicity, broad host range of infectivity, including non-dividing cells, and potential site-specific chromosomal integration make AAV an attractive tool for gene transfer.

[0002] Recent studies suggest that AAV vectors may be the preferred vehicle for gene therapy. To date, there have been 6 different serotypes of AAVs isolated from human or non-human primates (NHP) and well characterized. Among them, human serotype 2 is the first AAV that was developed as a gene transfer vector; it has been widely used for efficient gene transfer experiments in different target tissues and animal models. Gene therapy vectors based on adeno-associated virus type 1 have also been disclosed (Xiao et al. J. Virology; May 1999; pages 3994-4008). Clinical trials of the experimental application of AAV2 based vectors to some human disease models are in progress, and include such diseases as cystic fibrosis and hemophilia B.

[0003] A general PCR method suitable for detecting human papillomavirus types in cutaneous tumours and normal skin is known (Forslund et al J. of General Virology: 1999 80: P2437-2443).

[0004] What are desirable are AAV-based constructs for gene delivery.

#### SUMMARY OF THE INVENTION

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**[0005]** In one aspect, the invention provides a novel method of identifying unknown AAV sequences from cellular DNAs of various human and non-human primate (NHP) tissues using bioinformatics analysis, PCR based gene amplification and cloning technology, based on the nature of latency and integration of AAVs in the absence of helper virus co-infection, the method being defined in claim 1 hereinafter.

[0006] In another aspect the invention provides a kit for use in the method of the invention, the kit being as defined in claim 23 hereinafter.

#### DETAILED DESCRIPTION OF THE INVENTION

[0007] In the present invention, the inventors have found a method which takes advantage of the ability of adeno-associated virus (AAV) to penetrate the nucleus, and, in the absence of a helper virus co-infection, to integrate into cellular DNA and establish a latent infection. This method utilizes a polymerase chain reaction (PCR)-based strategy for detection, identification of sequences of AAVs from DNAs from tissues of human and non-human primate origin as well as from other sources.

40 [0008] Nucleic acid sequences can be identified according to the method of the invention. One such adeno-associated virus is of the serotype, termed herein serotype 7 (AAV7). Other novel adeno-associated virus serotypes identified by the method include AAV10, AAV11, and AAV12.

[0009] Among particularly desirable AAV fragments which can be identified are the cap proteins, including the vp1, vp2, vp3, the hypervariable regions, the rep proteins, including rep 78, rep 68, rep 52, and rep 40, and the sequences encoding these proteins. Each of these fragments may be readily utilized in a variety of vector systems and host cells. Such fragments may be used alone, in combination with other AAV sequences or fragments, or in combination with elements from other AAV or non-AAV viral sequences. In one particularly desirable embodiment, a vector contains the AAV cap and/or rep sequences.

[0010] As described herein, alignments are performed using any of a variety of publicly or commercially available Multiple Sequence Alignment Programs, such as "Clustal W", accessible through Web Servers on the internet. Alternatively, Vector NTI utilities are also used. There are also a number of algorithms known in the art which can be used to measure nucleotide sequence identity, including those contained in the programs described above. As another example, polynucleotide sequences can be compared using Fasta, a program in GCG Version 6.1. Fasta provides alignments and percent sequence identity of the regions of the best overlap between the query and search sequences. For instance, percent sequence identity between nucleic acid sequences can be determined using Fasta with its default parameters (a word size of 6 and the NOPAM factor for the scoring matrix) as provided in GCG Version 6.1. Similar programs are available for amino acid sequences, e.g., the "Clustal X" program. Generally, any of these programs are used at default settings, although one of skill in the art can alter these settings as needed. Alternatively, one of skill in the art can utilize

another algorithm or computer program which provides at least the level of identity or alignment as that provided by the referenced algorithms and programs.

**[0011]** The term "substantial homology" or "substantial similarity," when referring to a nucleic acid, or fragment thereof, indicates that, when optimally aligned with appropriate nucleotide insertions or deletions with another nucleic acid (or its complementary strand), there is nucleotide sequence identity in at least about 95 to 99% of the aligned sequences. Preferably, the homology is over full-length sequence, or an open reading frame thereof, or another suitable fragment which is at least 15 nucleotides in length. Examples of suitable fragments are described herein.

[0012] The term "substantial homology" or "substantial similarity," when referring to amino acids or fragments thereof, indicates that, when optimally aligned with appropriate amino acid insertions or deletions with another amino acid, there is amino acid sequence identity in at least about 95 to 99% of the aligned sequences. Preferably, the homology is over full-length sequence, or a protein thereof, e.g., a cap protein, a rep protein, or a fragment thereof which is at least 8 amino acids, or more desirably, at least 15 amino acids in length. Examples of suitable fragments are described herein.

[0013] By the term "highly conserved" is meant at least 80% identity, preferably at least 90% identity, and more preferably, over 97% identity. Identity is readily determined by one of skill in the art by resort to algorithms and computer programs known by those of skill in the art.

[0014] The term "percent sequence identity" or "identical" in the context of nucleic acid sequences refers to the residues in the two sequences which are the same when aligned for maximum correspondence. The length of sequence identity comparison may be over the full-length of the genome, the full-length of a gene coding sequence, or a fragment of at least about 500 to 5000 nucleotides, is desired. However, identity among smaller fragments, e.g. of at least about nine nucleotides, usually at least about 20 to 24 nucleotides, at least about 28 to 32 nucleotides, at least about 36 or more nucleotides, may also be desired. Similarly, "percent sequence identity" may be readily determined for amino acid sequences, over the full-length of a protein, or a fragment thereof. Suitably, a fragment is at least about 8 amino acids in length, and may be up to about 700 amino acids. Examples of suitable fragments are described herein.

[0015] The AAV sequences and fragments thereof are useful in production of rAAV, and are also useful as antisense delivery vectors, gene therapy vectors, or vaccine vectors.

[0016] As described herein, the vectors containing the AAV capsid proteins are particularly well suited for use in applications in which the neutralizing antibodies diminish the effectiveness of other AAV serotype based vectors, as well as other viral vectors. The rAAV vectors are particularly advantageous in rAAV readministration and repeat gene therapy.

[0017] As used throughout this specification and the claims, the terms "comprising" and "including" and their variants are inclusive of other components, elements, integers, steps and the like. Conversely, the term "consisting" and its variants is exclusive of other components, elements, integers, steps and the like.

#### I. Methods of the Invention

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## A. Detection of Sequences Via Molecular Cloning

**[0018]** In one aspect, the invention provides a method of identifying target (unknown) nucleic acid sequences in a sample. This method is particularly well suited for detection of viral sequences which are integrated into the chromosome of a cell, e.g., adeno-associated viruses (AAV) and retroviruses, among others.

[0019] As used herein, a sample is any source containing nucleic acids, e.g., tissue, tissue culture, cells, cell culture, and biological fluids including, without limitation, urine and blood. These nucleic acid sequences may be DNA or RNA from plasmids, natural DNA or RNA from any source, including bacteria, yeast, viruses, and higher organisms such as plants or animals. DNA or RNA is extracted from the sample by a variety of techniques known to those of skill in the art, such as those described by Sambrook, Molecular Cloning: A Laboratory Manual (New York: Cold Spring Harbor Laboratory). The origin of the sample and the method by which the nucleic acids are obtained for application of the method of the invention is not a limitation of the present invention. Optionally, the method of the invention can be performed directly on the source of DNA, or on nucleic acids obtained (e.g., extracted) from a source.

**[0020]** The method of the invention involves subjecting a sample containing DNA to amplification via polymerase chain reaction (PCR) using a first set of primers specific for a first region of double-stranded nucleic acid sequences, thereby obtaining amplified sequences.

[0021] As used herein, each of the "regions" is predetermined based upon the alignment of the nucleic acid sequences of at least two serotypes (e.g., AAV) or strains (e.g., lentiviruses), and wherein each of said regions is composed of sequences having a 5' end which is highly conserved, a middle which is variable, and a 3' end which is highly conserved, each of these being conserved or variable relative to the sequences of at least AAV1-AAV6. The 5' and 3' ends are highly conserved over at least 18 base pairs (bp). However, one or both of the sequences at the 5' or 3' end may be conserved over more than 18 bp, more than 25 bp, more than 30 bp, or more than 50 bp at the 5' end. With respect to the variable region, there is no requirement for conserved sequences, these sequences may be relatively conserved, or may have less than 90, 80, or 70% identity among the aligned serotypes or strains.

[0022] Each of the regions may span about 100 bp to about 10 kilobase pairs in length, provided that the first region is at least 250 bp in length. However, it is particularly desirable that one of the regions is a "signature region", i.e., a region which is sufficiently unique to positively identify the amplified sequence as being from the target source. For example, in one embodiment, the first region is about 250 bp in length, and is sufficiently unique among known AAV sequences, that it positively identifies the amplified region as being of AAV origin. Further, the variable sequences within this region are sufficiently unique that can be used to identify the serotype from which the amplified sequences originate. Once amplified (and thereby detected), the sequences can be identified by performing conventional restriction digestion and comparison to restriction digestion patterns for this region in any of AAV1, AAV2, AAV3, AAV4, AAV5, or AAV6, or that of AAV7, AAV10, AAV11, AAV12, or any of the other novel serotypes identified by the invention, which is predetermined and provided by the present invention.

**[0023]** Given the guidance provided herein, one of skill in the art can readily identify such regions among other integrated viruses to permit ready detection and identification of these sequences. Thereafter, an optimal set of generic primers located within the highly conserved ends can be designed and tested for efficient amplification of the selected region from samples. This aspect of the invention is readily adapted to a diagnostic kit for detecting the presence of the target sequence (e.g., AAV) and for identifying the AAV serotype, using standards which include the restriction patterns for the AAV serotypes described herein or isolated using the techniques described herein. For example, quick identification or molecular serotyping of PCR products can be accomplished by digesting the PCR products and comparing restriction patterns.

[0024] Thus, in one embodiment, the "signature region" for AAV spans about bp 2800 to about 3200 of AAV 1 [SEQ ID NO:6], and corresponding base pairs in AAV 2, AAV3, AAV4, AAV5, and AAV6. More desirably, the region is about 250 bp, located within bp 2886 to about 3143 bp of AAV 1 [SEQ ID NO:6], and corresponding base pairs in AAV 2 [SEQ ID NO:7], AAV3 [SEQ ID NO8], and other AAV serotypes. To permit rapid detection of AAV in the sample, primers which specifically amplify this signature region are utilized. However, the present invention is not limited to the exact sequences identified herein for the AAV signature region, as one of skill in the art may readily alter this region to encompass a shorter fragment, or a larger fragment of this signature region.

**[0025]** The PCR primers are generated using techniques known to those of skill in the art. Each of the PCR primer sets is composed of a 5' primer and a 3' primer. See, e.g., Sambrook et al, cited herein. The term "primer" refers to an oligonucleotide which acts as a point of initiation of synthesis when placed under conditions in which synthesis of a primer extension product which is complementary to a nucleic acid strand is induced. The primer is preferably single stranded. However, if a double stranded primer is utilized, it is treated to separate its strands before being used to prepare extension products. The primers may be about 15 to 25 or more nucleotides, and preferably at least 18 nucleotides. However, for certain applications shorter nucleotides, e.g., 7 to 15 nucleotides are utilized.

**[0026]** The primers are selected to be sufficiently complementary to the different strands of each specific sequence to be amplified to hybridize with their respective strands. Therefore, the primer sequence need not reflect the exact sequence of the region being amplified. For example, a non-complementary nucleotide fragment may be attached to the 5' end of the primer, with the remainder of the primer sequence being completely complementary to the strand. Alternatively, non-complementary bases or longer sequences can be interspersed into the primer, provided that the primer sequence has sufficient complementarity with the sequence of the strand to be amplified to hybridize therewith and form a template for synthesis of the extension product of the other primer.

[0027] The PCR primers for the signature region are based upon the highly conserved sequences of two or more aligned sequences (e.g., two or more AAV serotypes). The primers can accommodate less than exact identity among the two or more aligned AAV serotypes at the 5' end or in the middle. However, the sequences at the 3' end of the primers correspond to a region of two or more aligned AAV serotypes in which there is exact identity over at least five, preferably, over at least nine base pairs, and more preferably, over at least 18 base pairs at the 3' end of the primers. Thus, the 3' end of the primers is composed of sequences with 100% identity to the aligned sequences over at least five nucleotides. However, one can optionally utilize one, two, or more degenerate nucleotides at the 3' end of the primer. [0028] For example, the primer set for the signature region of AAV was designed based upon a unique region within the AAV capsid, as follows. The 5' primer was based upon nt 2867-2891 of AAV2 [SEQ ID NO:7], 5'-GGTAATTCCTCCGGAAATTGGCATT3'. The 3' primer was designed based upon nt 3096-3122 of AAV2 [SEQ ID NO:7], 5'-GACTCATCAACAACAACTGGGGATTC-'3. However, one of skill in the art may have readily designed the primer set based upon the corresponding regions of AAV 1, AAV3, AAV4, AAV5, AAV6, or based upon the information provided herein, AAV7. AAV10, AAV11, AAV12, or another novel AAV. In addition, still other primer sets can be readily designed to amplify this signature region, using techniques known to those of skill in the art.

#### B. Isolation of Target Sequences

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[0029] As described herein, the present invention uses a first primer set which specifically amplifies the signature region of the target sequence, e.g., an AAV serotype, in order to permit detection of the target. In a situation in which

further sequences are desired, e.g., if a novel AA V serotype is identified, the signature region may be extended. Thus, the invention may further utilize one or more additional primer sets.

**[0030]** Suitably, these primer sets are designed to include either the 5' or 3' primer of the first primer set and a second primer unique to the primer set, such that the primer set amplifies a region 5' or 3' to the signature region which anneals to either the 5' end or the 3' end of the signature region. For example, a first primer set is composed of a 5' primer, P1 and a 3' primer P2 to amplify the signature region. In order to extend the signature region on its 3' end, a second primer set is composed of primer P1 and a 3' primer P4, which amplifies the signature region and contiguous sequences downstream of the signature region. In order to extend the signature region on its 5' end, a third primer set is composed of a 5' primer, P5, and primer P2, such that the signature region and contiguous sequences upstream of the signature region are amplified. These extension steps are repeated (or performed at the same time), as needed or desired. Thereafter, the products results from these amplification steps are fused using conventional steps to produce an isolated sequence of the desired length.

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[0031] The second and third primer sets are designed, as with the primer set for the signature region, to amplify a region having highly conserved sequences among the aligned sequences. Reference herein to the term "second" or "third" primer set is for each of discussion only, and without regard to the order in which these primers are added to the reaction mixture, or used for amplification. The region amplified by the second primer set is selected so that upon amplification it anneals at its 5' end to the 3' end of the signature region. Similarly, the region amplified by the third primer set is selected so that upon amplification it anneals at its 3' end anneals to the 5' end of the signature region. Additional primer sets can be designed such that the regions which they amplify anneal to the either the 5' end or the 3' end of the extension products formed by the second or third primer sets, or by subsequent primer sets.

[0032] For example, where AAV is the target sequence, a first set of primers (P1 and P2) are used to amplify the signature region from the sample. In one desirable embodiment, this signature region is located within the AAV capsid. A second set of primers (P1 and P4) is used to extend the 3' end of the signature region to a location in the AAV sequence which is just before the AAV 3' ITR, i.e., providing an extension product containing the entire 3' end of the AAV capsid when using the signature region as an anchor. In one embodiment, the P4 primer corresponds to nt 4435 to 4462 of AAV2 [SEQ ID NO:7], and corresponding sequences in the other AAV serotypes. This results in amplification of a region of about 1.6 kb, which contains the 0.25 kb signature region. A third set of primers (P3 and P2) is used to extend the 5' end of signature region to a location in the AAV sequences which is in the 3' end of the rep genes, i.e., providing an extension product containing the entire 5' end of the AAV capsid when using the signature region as an anchor. In one embodiment, the P3 primer corresponds to nt 1384 to 1409 of AAV2 [SEQ ID NO:7], and corresponding sequences in the other AAV serotypes. This results in amplification of a region of about 1.7 kb, which contains the 0.25 kb signature region. Optionally, a fourth set of primers are used to further extend the extension product containing the entire 5' end of the AAV capsid to also include the rep sequences. In one embodiment, the primer designated P5 corresponds to nt 108 to 133 of AAV2 [SEQ ID NO:7], and corresponding sequences in the other AAV serotypes and is used in conjunction with the P2 primer.

**[0033]** Following completion of the desired number of extension steps, the various extension products are fused, making use of the signature region as an anchor or marker, to construct an intact sequence. In the example provided herein, AAV sequences containing, at a minimum, an intact AAV cap gene are obtained. Larger sequences may be obtained, depending upon the number of extension steps performed.

[0034] Suitably, the extension products are assembled into an intact AAV sequence using methods known to those of skill in the art. For example, the extension products may be digested with DrallI, which cleaves at the DrallI site located within the signature region, to provide restriction fragments which are re-ligated to provide products containing (at a minimum) an intact AAV cap gene. However, other suitable techniques for assembling the extension products into an intact sequence may be utilized. See, generally, Sambrook et al, cited herein.

[0035] As an alternative to the multiple extension steps described above, another embodiment of the invention provides for direct amplification of a 3.1 kb fragment which allows isolation of full-length cap sequences. To directly amplify a 3.1 kb full-length cap fragment from NHP tissue and blood DNAs, two other highly conserved regions were identified in AAV genomes for use in PCR amplification of large fragments. A primer within a conserved region located in the middle of the rep gene is utilized (AV1ns: 5' GCTGCGTCAACTGGACCAATGAGAAC 3', nt of SEQ ID NO:6) in combination with the 3' primer located in another conserved region downstream of the Cap gene (AV2cas: 5' CGCAGAGACCAAAGTTCAACTGAAACGA 3', SEQ ID NO:7) for amplification of AAV sequences including the full-length AAV cap. Typically, following amplification, the products are cloned and sequence analysis is performed with an accuracy of ≥ 99.9%. Using this method, the inventors have isolated at least 50 capsid clones which have subsequently been characterized. Among them, 37 clones were derived from Rhesus macaque tissues (rh.1 - rh.37), 6 clones from cynomologous macaques (cy.1 - cy.6), 2 clones from Baboons (bb.1 and bb.2) and 5 clones from Chimps (ch.1 - ch.5). These clones are identified elsewhere in the specification, together with the species of animal from which they were identified and the tissues in that animal these novel sequences have been located.

#### II. Diagnostic Kit

[0036] In another aspect, the invention provides a diagnostic kit as defined in claim 23 hereinafter for detecting the presence of an unknown adeno-associated virus (AAV) in a sample. Such a kit may contain a first set of 5' and 3' PCR primers specific for a signature region of the AAV nucleic acid sequence. Alternatively, or additionally, such a kit can contain a first set of 5' and 3' PCR primers specific for the 3.1 kb fragment which includes the full-length AAV capsid nucleic acid sequence identified herein (e.g., the AV1ns and AV2cas primers.) Optionally, a kit of the invention may further contain two or more additional sets of 5' and 3' primers, as described herein, and/or PCR probes. These primers and probes are used according to the present invention to amplify signature regions of each AAV serotype, e.g., using quantitative PCR.

[0037] Such a kit may further include one or more restriction enzymes, standards for AAV serotypes providing their "signature restriction enzyme digestions analyses", and/or other means for determining the serotype of the AAV detected. [0038] In addition, kits of the invention may include, instructions, a negative and/or positive control, containers, diluents and buffers for the sample, indicator charts for signature comparisons, disposable gloves, decontamination instructions, applicator sticks or containers, and sample preparator cups, as well as any desired reagents, including media, wash reagents and concentration reagents. Such reagents may be readily selected from among the reagents described herein, and from among conventional concentration reagents. In one desirable embodiment, the wash reagent is an isotonic saline solution which has been buffered to physiologic pH, such as phosphate buffered saline (PBS); the elution reagent is PBS containing 0.4 M NaCl, and the concentration reagents and devices. For example, one of skill in the art will recognize that reagents such as polyethylene glycol (PEG), or NH<sub>4</sub>SO<sub>4</sub> may be useful, or that devices such as filter devices. For example, a filter device with a 100 K membrane would concentrate rAAV.

[0039] The kits provided by the present invention are useful for performing the methods described herein, and for study of biodistribution, epidemiology, mode of transmission of novel AAV serotypes in human and NHPs.

[0040] Thus, the methods and kits of the invention permit identification of target AAV sequences, particularly integrated AAV sequences.

[0041] In one notable example, the method of the invention facilitated analysis of cloned AAV sequences by the inventors, which revealed heterogeneity of proviral sequences between cloned fragments from different animals, all of which were distinct from the known six AAV serotypes, with the majority of the variation localized to hypervariable regions of the capsid protein. Surprising divergence of AAV sequences was noted in clones isolated from single tissue sources, such as lymph node, from an individual rhesus monkey. This heterogeneity is best explained by apparent evolution of AAV sequence within individual animals due, in part, to extensive homologous recombination between a limited number of co-infecting parenteral viruses. These studies suggest sequence evolution of widely disseminated virus during the course of a natural AAV infection that presumably leads to the formation of swarms of quasispecies which differ from one another in the array of capsid hypervariable regions. This is the first example of rapid molecular evolution of a DNA virus in a way that formerly was thought to be restricted to RNA viruses.

[0042] Sequences of several novel AAV serotypes identified by the method of the invention and characterization of these serotypes is provided.

III. Novel AAV Serotypes

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#### A. Nucleic Acid Sequences

[0043] Nucleic acid sequences of novel AAV serotypes identified by the methods of the invention are provided. See, SEQ ID NO:1, 9 - 59, and 117 - 120. See also and the sequence listing.

<sup>45</sup> **[0044]** For novel serotype AAV7, the full-length sequences, including the AAV 5' ITRs, capsid, rep, and AAV 3' ITRs are provided in SEQ ID NO:1.

**[0045]** For other novel AA V serotypes, the approximately 3.1 kb fragment isolated according to the method of the invention is provided. This fragment contains sequences encoding full-length capsid protein and all or part of the sequences encoding the rep protein. These sequences include the clones identified below.

[0046] For still other novel AAV serotypes, the signature region encoding the capsid protein is provided. For example, the AAV10 nucleic acid sequences include those illustrated in See, SEQ ID NO:117, which spans 255 bases. The AAV11 nucleic acid sequences include the DNA sequences illustrated in SEQ ID NO:118 which spans 258 bases. The AAV12 nucleic acid sequences include the DNA sequences illustrated in SEQ ID NO: 119, which consists of 255 bases. Using the methodology described above, further AAV10, AAV11 and AAV 12 sequences can be readily identified and used for a variety of purposes, including those described for AAV7 and the other novel serotypes herein.

[0047] Novel NHP sequences identified by the invention include those provided in the following Table I, which are identified by clone number:

# Table 1

AAV Cap Sequence	Clone Number	Source		
		Species	Tissue	SEQ ID NO (DNA)
Rh.1	Clone 9 (AAV9)	Rhesus	Heart	5
Rh.2	Clone 43.1	Rhesus	MLN	39
Rh.3	Clone 43.5	Rhesus	MLN	40
Rh.4	Clone 43.12	Rhesus	MLN	41
Rh.5	Clone 43.20	Rhesus	MLN	42
Rh.6	Clone 43.21	Rhesus	MLN	43
Rh.7	Clone 43.23	Rhesus	MLN	44

# Table 1 (cont'd)

Rh.8	Clone 43.25	Rhesus	MLN	45
Rh.9	Clone 44.1	Rhesus	Liver	46
Rh.10	Clone 44.2	Rhesus	Liver	59
Rh.11	Clone 44.5	Rhesus	Liver	47
Rh.12	Clone	Rhesus	MLN	30
	42.1B			
Rh.13	42.2	Rhesus	MLN	9
Rh.14	Clone	Rhesus	MLN	32
· · · · · · · · · · · · · · · · · · ·	42.3A			<u> </u>
Rh.15	Clone	Rhesus	MLN	36
	42.3B			
Rh.16	Clone 42.4	Rhesus	MLN	33
Rh.17	Clone	Rhesus	MLN	34
	42.5A			
Rh.18	Clone	Rhesus	MLN	29
	42.5B	~		
Rh.19	Clone	Rhesus	MLN	38
7: 00	42.6B		N 41 N 1	27
Rh.20	Clone 42.8	Rhesus	MLN	27
Rh.21	Clone 42.10	Rhesus	MLN	35
Rh.22	Clone 42.11	Rhesus	MLN	37
Rh.23	Clone 42.12	Rhesus	MLN	58
Rh.24	Clone 42.13	Rhesus	MLN	31
Rh.25	Clone 42.15	Rhesus	MLN	28
Rh.26	Clone 223.2	Rhesus	Liver	49
Rh.27	Clone 223.4	Rhesus	Liver	50
Rh.28	Clone 223.5	Rhesus	Liver	51
Rh.29	Clone 223.6	Rhesus	Liver	52
Rh.30	Clone 223.7	Rhesus	Liver	53
Rh.31	Clone	Rhesus	Liver	48
D1: 32	223.10	Dhasus	Culon Due	10
Rh.32	Clone C1	Rhesus	Spleen, Duo, Kid & Liver	19
Rh.33	Clone C3	Rhesus	Kid & Liver	20
Rh.34	Clone C5	Rhesus		21
Rh.35	Clone F1	Rhesus	Liver	22
			Livei	23
Rh.36	Clone F3	Rhesus		24
Rh.37	Clone F5	Rhesus	Pland	<del></del>
Cy.1	Clone 1.3	Cyno	Blood	14
Cy.2	Clone 13.3B	Cyno	Blood	15
Cy.3	Clone 24.1	Cyno	Blood	16
Cy.4	Clone 27.3	Cyno	Blood	17
Cy.5	Clone 7.2	Cyno	Blood	18
Cy.6	Clone 16.3	Cyno	Blood	10
		·		<del> </del>

	Table 1 (cont'd)								
bb.1	Clone 29.3	Baboon	Blood	i i					
bb.2	Clone 29.5	Baboon	Blood	13					
Ch.1	Clone A3.3	Chimp	Blood	57					
Ch.2	Clone A3.4	Chimp	Blood	54					
Ch.3	Clone A3.5	Chimp	Blood	55					
Ch.4	Clone A3.7	Chimp	Blood	56					

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**[0048]** A novel NHP clone was made by splicing capsids fragments of two chimp adenoviruses into an AAV2 rep construct. This new clone, A3.1, is also termed Ch.5 [SEQ ID NO:20]. Additionally, the present invention includes two human AAV sequences, termed H6 [SEQ ID NO:25] and H2 [SEQ ID NO:26].

[0049] The AAV nucleic acid sequences further encompass the strand which is complementary to the strands provided in the sequences provided in the Sequence Listing [SEQ ID NO:1, 9 - 59, 117-120], nucleic acid sequences, as well as the RNA and cDNA sequences corresponding to the sequences provided in the Sequence Listing [SEQ ID NO:1, 9-59, 117-120], and their complementary strands. Also included in the nucleic acid sequences are natural variants and engineered modifications of the sequences of the Sequence Listing [SEQ ID NO:1, 9 - 59, 117-120], and their complementary strands. Such modifications include, for example, labels which are known in the art, methylation, and substitution of one or more of the naturally occurring nucleotides with a degenerate nucleotide.

**[0050]** Further included are nucleic acid sequences which are greater than 85%, preferably at least about 90%, more preferably at least about 95%, and most preferably at least about 98 to 99% identical or homologous to the sequences of the invention, including the Sequence Listing [SEQ ID NO:1, 9 - 59, 117-120]. These terms are as defined herein.

**[0051]** Also included are fragments of the novel AAV sequences identified by the method described herein. Suitable fragments are at least 15 nucleotides in length, and encompass functional fragments, i.e., fragments which are of biological interest. In one embodiment, these fragments are fragments of the novel sequences of the Sequence Listing [SEQ ID NO:1, 9 - 59, 117-120], their complementary strands, cDNA and RNA complementary thereto.

**[0052]** Examples of suitable fragments are provided with respect to the location of these fragments on AAV1, AAV2, or AAV7. However, using the alignment provided herein (obtained using the Clustal W program at default settings), or similar techniques for generating an alignment with other novel serotypes of the invention, one of skill in the art can readily identify the precise nucleotide start and stop codons for desired fragments.

[0053] Examples of suitable fragments include the sequences encoding the three variable proteins (vp) of the AAV capsid which are alternative splice variants: vp1 [e.g., nt 825 to 3049 of AA V7, SEQ ID NO: 1]; vp2 [e.g., nt 1234 - 3049 of AAV7, SEQ ID NO: 1]; and vp 3 [e.g., nt 1434 - 3049 of AAV7, SEQ ID NO:1]. It is notable that AAV7 has an unusual GTG start codon. With the exception of a few house-keeping genes, such a start codon has not previously been reported in DNA viruses. The start codons for vp1, vp2 and vp3 for other AAV serotypes have been believed to be such that they permit the cellular mechanism of the host cell in which they reside to produce vp1, vp2 and vp3 in a ratio of 10%:10%:80%, respectively, in order to permit efficient assembly of the virion. However, the AAV7 virion has been found to assemble efficiently even with this rare GTG start codon. Thus, the inventors anticipate this it is desirable to alter the start codon of the vp3 of other AAV serotypes to contain this rare GTG start codon, in order to improve packaging efficiency, to alter the virion structure and/or to alter location of epitopes (e.g., neutralizing antibody epitopes) of other AAV serotypes. The start codons may be altered using conventional techniques including, e.g., site directed mutagenesis. The altered AAV virions may be of any selected serotype, composed of a vp 3, and/or optionally, vp 1 and/or vp2 having start codons altered to GTG.

[0054] Other suitable fragments of AAV, include a fragment containing the start codon for the AAV capsid protein [e.g., nt 468 to 3090 of AAV7, SEQ ID NO:1, nt 725 to 3090 of AAV7, SEQ ID NO:1, and corresponding regions of the other AAV serotypes]. Still other fragments of AAV7 and the other novel AAV semtypes identified using the methods described herein include those encoding the rep proteins, including *rep* 78 [e.g., initiation codon 334 for AAV7], *rep* 68 [initiation codon nt 334 for AAV7], *rep* 52 [initiation codon 1006 for AAV7], and *rep* 40 [initiation codon 1006 for AAV7] Other fragments of interest may include the AAV 5' inverted terminal repeats ITRs, [nt 1 to 107 for AAV7]; the AA V 3' ITRs [nt 4704 to 4721 for AAV7], P19 sequences. AAV P40 sequences, the rep binding site, and the terminal resolute site (TRS). Still other suitable fragments wilt be readily apparent to those of skill in the art.

[0055] In addition to the nucleic acid sequences provided in the figures and Sequence Listing, there are nucleic acid molecules and sequences which are designed to express the amino acid sequences, proteins and peptides of the AAV serotypes of the invention. These include nucleic acid sequences which encode the following novel AAV amino acid sequences: C1 [SEQ ID NO:60], C2 [SEQ ID NO:61], C5 [SEQ ID NO:62], A3-3 [SEQ ID NO:66], A3-7 [SEQ ID NO:67],

A3-4 [SEQ ID NO:68], A3-5 [SEQ ID NO: 69], 3.3b [SEQ ID NO: 62], 223.4 [SEQ ID NO: 73], 223-5 [SEQ ID NO:74], 223-10 [SEQ ID NO:75], 223-2 [SEQ ID NO:76], 223-7 [SEQ ID NO: 77], 223-6 [SEQ ID NO: 78], 44-1 [SEQ ID NO: 79], 44-5 [SEQ ID NO:80], 44-2 [SEQ ID NO:81], 42-15 [SEQ ID NO: 84], 42-8 [SEQ ID NO: 85], 42-13 [SEQ ID NO:86], 42-3A [SEQ ID NO:87], 42-4 [SEQ ID NO:88], 42-5A [SEQ ID NO:89], 42-1B [SEQ ID NO:90], 42-5B [SEQ ID NO:91], 43-1 [SEQ ID NO: 92], 43-12 [SEQ ID NO: 93], 43-5 [SEQ ID NO:94], 43-21 [SEQ ID NO:96], 43-25 [SEQ ID NO: 97], 43-20 [SEQ ID NO:99], 24.1 [SEQ ID NO: 101], 42.2 [SEQ ID NO:102], 7.2 [SEQ ID NO: 103], 27.3 [SEQ ID NO: 104], 16.3 [SEQ ID NO: 105], 42.10 [SEQ ID NO: 106], 42-38 [SEQ ID NO: 107], 42-11 [SEQ ID NO: 108], F1 [SEQ ID NO: 109], F5 [SEQ ID NO: 110], F3 [SEQ ID NO:111], 42-6B [SEQ ID NO: 112], and/or 42-12 [SEQ ID NO: 113], and artificial AAV serotypes generated using these sequences and/or unique fragments thereof.

**[0056]** As used herein, artificial AAV serotypes include, without limitation. AAV with a non-naturally occurring capsid protein. Such an artificial capsid may be generated by any suitable technique, using a novel AAV sequence (e.g., a fragment of a vp1 capsid protein) in combination with heterologous sequences which may be obtained from another AAV serotype (known or novel), non-contiguous portions of the same AAV serotype, from a non-AAV viral source, or from a non-viral source. An artificial AAV serotype may be, without limitation, a chimeric AAV capsid, a recombinant AAV capsid, or a "humanized" AAV capsid.

#### B. AAV Amino Acid Sequences, Proteins and Peptides

[0057] The invention provides proteins and fragments thereof which are encoded by the nucleic acid sequences of the novel AAV serotypes identified herein, including, e.g., AA V7 [nt 825 to 3049 of AA V7, SEQ ID NO: 1] the other novel serotypes provided herein. Thus, the capsid proteins of the novel serotypes of the invention, including: H6 [SEQ ID NO: 25], H2 [SEQ ID NO: 26], 42-2 [SEQ ID NO:9], 42-8 [SEQ ID NO:27], 42-15 [SEQ ID NO:28], 42-5b [SEQ ID NO: 29], 42-1b [SEQ ID NO:30]; 42-13 [SEQ ID NO: 31], 42-3a [SEQ ID NO: 32], 42-4 [SEQ ID NO:33], 42-5a [SEQ ID NO: 34], 42-10 [SEQ ID NO:35], 42-3b [SEQ ID NO: 36], 42-11 [SEQ ID NO: 37], 42-6b [SEQ ID NO:38], 43-1 [SEQ ID NO: 39], 43-5 [SEQ ID NO: 40], 43-12 [SEQ ID NO:41], 43-20 [SEQ ID NO:42], 43-21 [SEQ ID NO: 43], 43-23 [SEQ ID NO:44], 43-25 [SEQ ID NO: 45], 44.1 [SEQ ID NO:47], 44.5 [SEQ ID NO:47], 223.10 [SEQ ID NO:48], 223.2 [SEQ ID NO:49], 223.4 [SEQ ID NO:50], 223.5 [SEQ ID NO:51], 223.6 [SEQ ID NO:57], 42.12 [SEQ ID NO: 58], A3.4 [SEQ ID NO: 54], A3.5 [SEQ ID NO:55], A3.7 [SEQ ID NO: 56], A3.3 [SEQ ID NO:57], 42.12 [SEQ ID NO: 58], and 44.2 [SEQ ID NO: 59], can be readily generated using conventional techniques from the open reading frames provided for the above-listed clones.

**[0058]** The sequences, proteins, and fragments may be produced by any suitable means, including recombinant production, chemical synthesis, or other synthetic means. Such production methods are within the knowledge of those of skill in the art.

#### 35 IV. Production of rAAV with novel AAV capsids

**[0059]** Novel, wild-type AAV serotypes can be identified by the invention, the sequences of which wild-type AAV serotypes are free of DNA and/or cellular material with these viruses are associated in nature. In another aspect, the present invention provides molecules which utilize the novel AAV sequences of the invention, including fragments thereof, for production of molecules useful in delivery of a heterologous gene or other nucleic acid sequences to a target cell. **[0060]** The following examples illustrate several aspects and embodiments of the invention.

#### **EXAMPLES**

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45 Example 1: PCR amplification, cloning and characterization of novel AAV sequences.

[0061] Tissues from nonhuman primates were screened for AAV sequences using a PCR method based on oligonucleotides to highly conserved regions of known AAVs. A stretch of AAV sequence spanning 2886 to 3143 bp of AAV1 [SEQ ID NO:6] was selected as a PCR amplicon in which a hypervariable region of the capsid protein (Cap) that is unique to each known AAV serotype, which is termed herein a "signature region," is flanked by conserved sequences. In later analysis, this signature region was shown to be located between conserved residues spanning hypervariable region 3.

[0062] An initial survey of peripheral blood of a number of nonhuman primate species revealed detectable AAV in a subset of animals from species such as rhesus macaques, cynomologous macaques, chimpanzees and baboons. However, there were no AAV sequences detected in some other species tested, including Japanese macaques, pig-tailed macaques and squirrel monkeys. A more extensive analysis of vector distribution was conducted in tissues of rhesus monkeys of the University of Pennsylvania and Tulane colonies recovered at necropsy. This revealed AAV sequence throughout a wide array of tissues.

#### A. Amplification of an AAV signature region

[0063] DNA sequences of AAV1-6 and AAVs isolated from Goose and Duck were aligned to each other using "Clustal W" at default settings. Sequence similarities among AAVs were compared.

[0064] In the line of study, a 257 bp region spanning 2886 bp to 3143 bp of AAV 1 [SEQID NO: 6], and the corresponding region in the genomes of AAV 2-6 genomes was identified by the inventors. This region is located with the AAV capsid gene and has highly conserved sequences among at both 5' and 3' ends and is relatively variable sequence in the middle. In addition, this region contains a DrallI restriction enzyme site (CACCACGTC, SEQ ID NO:15). The inventors have found that this region serves as specific signature for each known type of AAV DNA. In other words, following PCR reactions, digestion with endonucleases that are specific to each known serotypes and gel electrophoresis analysis, this regions can be used to definitively identify amplified DNA as being from serotype 1, 2, 3, 4, 5, 6, or another serotype. [0065] The primers were designed, validated and PCR conditions optimized with AAV1, 2 and 5 DNA controls. The primers were based upon the sequences of AAV2: 5' primer, 1S: bp 2867-2891 of AAV2 (SEQ ID NO:7) and 3' primer, 18as, bp 3095-3121 of AAV2 (SEQ ID NO:7).

**[0066]** Cellular DNAs from different tissues including blood, brain, liver, lung, testis, etc. of different rhesus monkeys were studied utilizing the strategy described above. The results revealed that DNAs from different tissues of these monkeys gave rise to strong PCR amplifications. Further restriction analyses of PCR products indicated that they were amplified from AAV sequences different from any published AAV sequences.

[0067] PCR products (about 255 bp in size) from DNAs of a variety of monkey tissues have been cloned and sequenced. Bioinformatics study of these novel AAV sequences indicated that they are novel AAV sequences of capsid gene and distinct from each other. Multiple sequence alignment analysis was performed using the Clustal W (1.81) program. The percentage of sequence identity between the signature regions of AAV 1-7 and AAV 10-12 genomes is provided below.

Table 1. Sequences for Analysis

Sequence #	AAV Serotype	Size (bp)
1	AAV1	258
2	AAV2	255
3	AAV3	255
4	AAV4	246
5	AAV5	258
6	AAV6	258
7	AAV7	258
10	AAV10	255
11	AAV11	258
12	AAV12	255

Table 3. Pairwise Alignment (Percentage of Identity)

	AAV2	AAV3	AAV4	AAV5	AAV6	AAV7	AAV10	AAV11	AAV12
AAV1	90	90	81	76	97	91	93	94	93
AAV2		93	79	78	90	90	93	93	92
AAV3			80	76	90	92	92	92	92
AAV4				76	81	84	82	81	79
AAV5					75	78	79	79	76
AAV6						91	92	94	94
AAV7							94	92	92
AAV10								95	93

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#### Table continued

	AAV2	AAV3	AAV4	AAV5	AAV6	AAV7	AAV10	AAV11	AAV12
AAV11									94

**[0068]** Over 300 clones containing novel AAV serotype sequences that span the selected 257 bp region were isolated and sequenced. Bioinformatics analysis of these 300+ clones suggests that this 257 bp region is critical in serving as a good land marker or signature sequence for quick isolation and identification of novel AAV serotype.

B. Use of the signature region for PCR amplification.

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**[0069]** The 257 bp signature region was used as a PCR anchor to extend PCR amplifications to 5' of the genome to cover the junction region of rep and cap genes (1398 bp - 3143 bp, SEQ ID NO:6) and 3' of the genome to obtain the entire cap gene sequence (2866 bp - 4600 bp, SEQ ID NO:6). PCR amplifications were carried out using the standard conditions, including denaturing at 95°C for 0.5-1 min, annealing at 60-65°C for 0.5-1 min and extension at 72° C for I min per kb with a total number of amplification cycles ranging from 28 to 42.

[0070] Using the aligned sequences as described in "A", two other relative conserved regions were identified in the sequence located in 3' end of rep genes and 5' to the 257 bp region and in the sequence down stream of the 257 bp fragment but before the AAV' 3 ITR. Two sets of new primers were designed and PCR conditions optimized for recovery of entire capsid and a part of rep sequences of novel AAV serotypes. More specifically, for the 5' amplification, the 5' primer, AV1Ns, was GCTGCGTCAACTGGACCAATGAGAAC [nt 1398-1423 of AAV1, SEQ ID NO:6] and the 3' primer was 18as, identified above. For the 3' amplification, the 5' primer was 1s, identified above, and the 3' primer was AV2Las, TCGTTTCAGTTGAACTTTGGTCTCTGCG [nt 4435-4462 of AAV2, SEQ ID NO:7].

[0071] In these PCR amplifications, the 257 bp region was used as a PCR anchor and land marker to generate overlapping fragments to construct a complete capsid gene by fusion at the DrallI site in the signature region following amplification of the 5' and 3' extension fragments obtained as described herein. More particularly, to generate the intact AAV7 cap gene, the three amplification products (a) the sequences of the signature region; (b) the sequences of the 5' extension; and (c) the sequences of the 3' extension were cloned into a pCR4-Topo [Invitrogen] plasmid backbone according to manufacturer's instructions. Thereafter, the plasmids were digested with DrallI and recombined to form an intact cap gene

[0072] In this line of work, about 80 % of capsid sequences of AAV7 and AAV 8 were isolated and analyzed. Another novel serotype, AAV9, was also discovered from Monkey #2.

**[0073]** Using the PCR conditions described above, the remaining portion of the rep gene sequence for AAV7 is isolated and cloned using the primers that amplify 108 bp to 1461 bp of AAV genome (calculated based on the numbering of AAV2, SEQ ID NO:7). This clone is sequenced for construction of a complete AAV7 genome without ITRs.

C. Direct Amplification of 3.1 kb Cap fragment

[0074] To directly amplify a 3.1 kb full-length Cap fragment from NHP tissue and blood DNAs, two other highly conserved regions were identified in AAV genomes for use in PCR amplification of large fragments. A primer within a conserved region located in the middle of the rep gene was selected (AV1ns: 5' GCTGCGTCAACTGGACCAATGAGAAC 3', nt 1398-1423 of SEQ ID NO:6) in combination with the 3' primer located in another conserved region downstream of the Cap gene (AV2cas: 5' CGCAGAGACCAAAGTTCAACTGAAACGA 3', SEQ ID NO:7) for amplification of full-length cap fragments. The PCR products were Topo-cloned according to manufacturer's directions (Invitrogen) and sequence analysis was performed by Qiagengenomics (Qiagengenomics, Seattle, WA) with an accuracy of ≥ 99.9%. A total of 50 capsid clones were isolated and characterized. Among them, 37 clones were derived from Rhesus macaque tissues (rh.1 - rh.37), 6 clones from cynomologous macaques (cy.1 - cy.6), 2 clones from Baboons (bb.1 and bb.2) and 5 clones from Chimps (ch.1 - ch.5).

[0075] To rule out the possibility that sequence diversity within the novel AAV family was not an artifact of the PCR, such as PCR-mediated gene splicing by overlap extension between different partial DNA templates with homologous sequences, or the result of recombination process in bacteria, a series of experiments were performed under identical conditions for VP1 amplification using total cellular DNAs. First, intact AAV7 and AAV8 plasmids were mixed at an equal molar ratio followed by serial dilutions. The serially diluted mixtures were used as templates for PCR amplification of 3.1 kb VP1 fragments using universal primers and identical PCR conditions to that were used for DNA amplifications to see whether any hybrid PCR products were generated. The mixture was transformed into bacteria and isolated transformants to look for hybrid clones possibly derived from recombination process in bacterial cells. In a different experiment, we restricted AAV7 and AAV8 plasmids with Msp I, Ava I and HaeI, all of which cut both genomes multiple times at different

positions, mixed the digestions in different combinations and used them for PCR amplification of VP1 fragments under the same conditions to test whether any PCR products could be generated through overlap sequence extension of partial AAV sequences. In another experiment, a mixture of gel purified 5' 1.5 kb AAV7 VP1 fragment and 3' 1.7 kb AAV8 VP1 fragment with overlap in the signature region was serially diluted and used for PCR amplification in the presence and absence of 200 ng cellular DNA extracted from a monkey cell line that was free of AAV sequences by TaqMan analysis. None of these experiments demonstrated efficient PCR-mediated overlap sequence production under the conditions of the genomic DNA Cap amplification (data not shown). As a further confirmation, 3 pairs of primers were designed, which were located at different HVRs, and were sequence specific to the variants of clone 42s from Rhesus macaque F953, in different combinations to amplify shorter fragments from mesenteric lymph node (MLN) DNA from F953 from which clone 42s were isolated. All sequence variations identified in full-length Cap clones were found in these short fragments (data not shown).

Example 2: Adeno-Associated Viruses Undergo Substantial Evolution in Primates During Natural Infections

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[0076] Sequence analysis of selected AAV isolates revealed divergence throughout the genome that is most concentrated in hypervariable regions of the capsid proteins. Epidemiologic data indicate that all known serotypes are endemic to primates, although isolation of clinical isolates has been restricted to AAV2 and AAV3 from an all and throat swabs of human infants and AAV5 from a human condylomatous wart. No known clinical sequalae have been associated with AAV infection.

[0077] In an attempt to better understand the biology of AAV, nonhuman primates were used as models to characterize the sequlae of natural infections. Tissues from nonhuman primates were screened for AAV sequences using the PCR method of the invention based on oligonucleotides to highly conserved regions of known AAVs (see Example 1). A stretch of AAV sequence spanning 2886 to 3143 bp of AAV1 [SEQ ID NO:6] was selected as a PCR amplicon in which conserved sequences are flanked by a hypervariable region that is unique to each known AAV serotype, termed herein a "signature region."

[0078] An initial survey of peripheral blood of a number of nonhuman primate species including rhesus monkeys, cynomologous monkeys, chimpanzees, and baboons revealed detectable AAV in a subset of animals from all species. A more extensive analysis of vector distribution was conducted in tissues of rhesus monkeys of the University of Pennsylvania and Tulane colonies recovered at necropsy. This revealed AAV sequence throughout a wide array of tissues. [0079] The amplified signature sequences were subcloned into plasmids and individual transformants were subjected to sequence analysis. This revealed substantial variation in nucleotide sequence of clones derived from different animals. Variation in the signature sequence was also noted in clones obtained within individual animals. Tissues harvested from two animals in which unique signature sequences were identified (i.e., colon from 98E044 and heart from 98E056) were further characterized by expanding the sequence amplified by PCR using oligonucleotides to highly conserved sequences. In this way, complete proviral structures were reconstructed for viral genomes from both tissues as described herein. These proviruses differ from the other known AAVs with the greatest sequence divergence noted in regions of the Cap gene.

[0080] Additional experiments were performed to confirm that AAV sequences resident to the nonhuman primate tissue represented proviral genomes of infectious virus that is capable of being rescued and form virions. Genomic DNA from liver tissue of animal 98E056, from which AAV8 signature sequence was detected, was digested with an endonuclease that does not have a site within the AAV sequence and transfected into 293 cells with a plasmid containing an E1 deleted genome of human adenovirus serotype 5 as a source of helper functions. The resulting lysate was passaged on 293 cells once and the lysate was recovered and analyzed for the presence of AAV Cap proteins using a broadly reacting polyclonal antibody to Cap proteins and for the presence and abundance of DNA sequences from the PCR amplified AAV provirus from which AAV8 was derived. Transfection of endonuclease restricted heart DNA and the adenovirus helper plasmid yielded high quantities of AAV8 virus as demonstrated by the detection of Cap proteins by Western blot analysis and the presence of 10<sup>4</sup> AAV8 vector genomes per 293 cell. Lysates were generated from a large-scale preparation and the AAV was purified by cesium sedimentation. The purified preparation demonstrated 26 nm icosohedral structures that look identical to those of AAV serotype 2. Transfection with the adenovirus helper alone did not yield AAV proteins or genomes, ruling out contamination as a source of the rescued AAV.

[0081] To further characterize the inter and intra animal variation of AAV signature sequence, selected tissues were subjected to extended PCR to amplify entire Cap open reading frames.

[0082] The resulting fragments were cloned into bacterial plasmids and individual transformants were isolated and fully sequenced. This analysis involved mesenteric lymph nodes from three rhesus monkeys (Tulane/V223 - 6 clones; Tulane/T612 - 7 clones; Tulane/F953 - 14 clones), liver from two rhesus monkeys (Tulane/V251 - 3 clones; Penn/00E033 - 3 clones), spleen from one rhesus monkey (Penn/97E043 - 3 clones), heart from one rhesus monkey (IHGT/98E046-1 clone) and peripheral blood from one chimpanzee (New Iberia/X133 - 5 clones), six cynomologous macaques (Charles River/A1378, A3099, A3388, A3442, A2821, A3242 - 6 clones total) and one Baboon (SFRB/8644 - 2 clones). Of the

50 clones that were sequenced from 15 different animals, 30 were considered non-redundant based on the finding of at least 7 amino acid differences from one another. The non-redundant VP1 clones are numbered sequentially as they were isolated, with a prefix indicating the species of non-human primate from which they were derived. The structural relationships between these 30 non-redundant clones and the previously described 8 AAV serotypes were determined using the SplitsTree program [Huson, D. H. SplitsTree: analyzing and visualizing evolutionary data. *Bioinformatics* 14, 68-73 (1998)] with implementation of the method of split decomposition. The analysis depicts homoplasy between a set of sequences in a tree-like network rather than a bifurcating tree. The advantage is to enable detection of groupings that are the result of convergence and to exhibit phylogenetic relationships even when they are distorted by parallel events. Extensive phylogenetic research will be required in order to elucidate the AAV evolution, whereas the intention here only is to group the different clones as to their sequence similarity.

**[0083]** To confirm that the novel VP1 sequences were derived from infectious viral genomes, cellular DNA from tissues with high abundance of viral DNA was restricted with an endonuclease that should not cleave within AAV and transfected into 293 cells, followed by infection with adenovirus. This resulted in rescue and amplification of AAV genomes from DNA of tissues from two different animals (data not shown).

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[0084] VP1 sequences of the novel AAVs were further characterized with respect to the nature and location of amino acid sequence variation. All 30 VP1 clones that were shown to differ from one another by greater than 1% amino acid sequence were aligned and scored for variation at each residue. An algorithm developed to determine areas of sequence divergence yielded 12 hypervariable regions (HVR) of which 5 overlap or are part of the 4 previously described variable regions [Kotin, cited above; Rutledge, cited above]. The threefold-proximal peaks contain most of the variability (HVR5-10). Interestingly the loops located at the 2 and 5 fold axis show intense variation as well. The HVRs 1 and 2 occur in the N-terminal portion of the capsid protein that is not resolved in the X-ray structure suggesting that the N-terminus of the VP1 protein is exposed on the surface of the virion.

[0085] Real-time PCR was used to quantify AAV sequences from tissues of 21 rhesus monkeys using primers and probes to highly conserved regions of Rep (one set) and Cap (two sets) of known AAVs. Each data point represents analysis from tissue DNA from an individual animal. This confirmed the wide distribution of AAV sequences, although the quantitative distribution differed between individual animals. The source of animals and previous history or treatments did not appear to influence distribution of AAV sequences in rhesus macaques. The three different sets of primers and probes used to quantify AAV yielded consistent results. The highest levels of AAV were found consistently in mesenteric lymph nodes at an average of 0.01 copies per diploid genome for 13 animals that were positive. Liver and spleen also contained high abundance of virus DNA. There were examples of very high AAV, such as in heart of rhesus macaque 98E056, spleen of rhesus macaque 97E043 and liver of rhesus macaque RQ4407, which demonstrated 1.5, 3 and 20 copies of AAV sequence per diploid genome respectively. Relatively low levels of virus DNA were noted in peripheral blood mononuclear cells, suggesting the data in tissue are not due to resident blood components (data not shown). It should be noted that this method would not necessarily capture all AAVs resident to the nonhuman primates since detection requires high homology to both the oligonucleotides and the real time PCR probe. Tissues from animals with high abundance AAV DNA was further analyzed for the molecular state of the DNA, by DNA hybridization techniques, and its cellular distribution, by *in situ* hybridization.

**[0086]** The kind of sequence variation revealed in AAV proviral fragments isolated from different animals and within tissues of the same animals is reminiscent of the evolution that occurs for many RNA viruses during pandemics or even within the infection of an individual. In some situations the notion of a wild-type virus has been replaced by the existence of swarms of quasispecies that evolve as a result of rapid replication and mutations in the presence of selective pressure. One example is infection by HIV, which evolves in response to immunologic and pharmacologic pressure. Several mechanisms contribute to the high rate of mutations in RNA viruses, including low fidelity and lack of proof reading capacity of reverse transcriptase and non-homologous and homologous recombination.

[0087] Evidence for the formation of quasispecies of AAV was illustrated in this study by the systematic sequencing of multiple cloned proviral fragments. In fact, identical sequences could not be found within any extended clones isolated between or within animals. An important mechanism for this evolution of sequence appears to be a high rate of homologous recombination between a more limited number of parenteral viruses. The net result is extensive swapping of hypervariable regions of the Cap protein leading to an array of chimeras that could have different tropisms and serologic specificities (i.e., the ability to escape immunologic responses especially as it relates to neutralizing antibodies). Mechanisms by which homologous recombination could occur are unclear. One possibility is that + and - strands of different single stranded AAV genomes anneal during replication as has been described during high multiplicity of infections with AAV recombinants. It is unclear if other mechanisms contribute to sequence evolution in AAV infections. The overall rate of mutation that occurs during AAV replication appears to be relatively low and the data do not suggest high frequencies of replication errors. However, substantial rearrangements of the AAV genome have been described during lytic infection leading to the formation of defective interfering particles. Irrespective of the mechanisms that lead to sequence divergence, with few exceptions, vp1 structures of the quasispecies remained intact without frameshifts or nonsense mutations suggesting that competitive selection of viruses with the most favorable profile of fitness contribute to the population

dynamics.

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**[0088]** These studies have implications in several areas of biology and medicine. The concept of rapid virus evolution, formerly thought to be a property restricted to RNA viruses, should be considered in DNA viruses, which classically have been characterized by serologic assays. It will be important in terms of parvoviruses to develop a new method for describing virus isolates that captures the complexity of its structure and biology, such as with HIV, which are categorized as general families of similar structure and function called Clades. An alternative strategy is to continue to categorize isolates with respect to serologic specificity and develop criteria for describing variants within serologic groups.

Example 3: Vectorology of recombinant AAV genomes equipped with AAV2 ITRs using chimeric plasmids containing AAV2 rep and novel AAV cap genes for scrological and gene transfer studies in different animal models.

[0089] Chimeric packaging constructs are generated by fusing AAV2 rep with cap sequences of novel AAV serotypes. These chimeric packaging constructs are used, initially, for pseudotyping recombinant AAV genomes carrying AAV2 ITRs by triple transfection in 293 cell using Ad5 helper plasmid. These pseudotyped vectors are used to evaluate performance in transduction-based serological studies and evaluate gene transfer efficiency of novel AAV serotypes in different animal models including NHP and rodents, before intact and infectious viruses of these novel serotypes are isolated.

A. pAAV2GFP

**[0090]** The AAV2 plasmid which contains the AAV2 ITRs and green fluorescent protein expressed under the control of a constitutitive promoter. This plasmid contains the following elements: the AAV2 ITRs, a CMV promoter, and the GFP coding sequences.

25 B. Cloning of trans plasmid

[0091] To construct the chimeric trans-plasmid for production of recombinant pseudotyped AAV7 vectors, p5E18 plasmid (Xiao *et al.*, 1999, *J. Virol* 73:3994-4003) was partially digested with Xho I to linearize the plasmid at the Xho I site at the position of 3169 bp only. The Xho I cut ends were then filled in and ligated back. This modified p5E18 plasmid was restricted with Xba I and Xho I in a complete digestion to remove the AAV2 cap gene sequence and replaced with a 2267 bp Spe I/Xho I fragment containing the AAV7 cap gene which was isolated from pCRAAV7 6-5+15-4 plasmid. [0092] The resulting plasmid contains the AAV2 rep sequences for Rep78/68 under the control of the AAV2 P5 promoter, and the AAV2 rep sequences for Rep52/40 under the control of the AAV2 P19 promoter. The AAV7 capsid sequences are under the control of the AAV2 P40 promoter, which is located within the Rep sequences. This plasmid further contains a spacer 5' of the rep ORF.

## C. Production of Pseudotyped rAAV

[0093] The rAAV particles (AAV2 vector in AAV7 capsid) are generated using an adenovirus-free method. Briefly, the cis plasmid (pAAV2.1 lacZ plasmid containing AAV2 ITRs), and the trans plasmid pCRAAV7 6-5+15-4 (containing the AAV2 rep and AAV7 cap) and a helper plasmid, respectively, were simultaneously co-transfected into 293 cells in a ratio of 1:1:2 by calcium phosphate precipitation.

[0094] For the construction of the pAd helper plasmids, pBG 10 plasmid was purchased from Microbix (Canada). A RsrII fragment containing L2 and L3 was deleted from pBHG10, resulting in the first helper plasmid, pAd $\Delta$ F13. Plasmid Ad $\Delta$ F1 was constructed by cloning Asp700/SalI fragment with a Pmel/SgfI deletion, isolating from pBHG10, into Bluescript. MLP, L2, L2 and L3 were deleted in the pAd $\Delta$ F1. Further deletions of a 2.3 kb NruI fragment and, subsequently, a 0.5 kb RsrII/NruI fragment generated helper plasmids pAd $\Delta$ F5 and pAd $\Delta$ F6, respectively. The helper plasmid, termed p $\Delta$ F6, provides the essential helper functions of E2a and E4 ORF6 not provided by the E1-expressing helper cell, but is deleted of adenoviral capsid proteins and functional E1 regions).

[0095] Typically, 50 μg of DNA (cis:trans:helper) was transfected onto a 150 mm tissue culture dish. The 293 cells were harvested 72 hours post-transfection, sonicated and treated with 0.5% sodium deoxycholate (37°C for 10 min.) Cell lysates were then subjected to two rounds of a CsCl gradient. Peak fractions containing rAAV vector are collected, pooled and dialyzed against PBS.

Example 4: Creation of infectious clones carrying intact novel AAV serotypes for study of basic virology in human and NHP derived cell lines and evaluation of pathogenesis of novel AAV serotypes in NHP and other animal models.

[0096] To achieve this goal, the genome walker system is employed to obtain 5' and 3' terminal sequences (ITRs)

and complete construction of clones containing intact novel AAV serotype genomes.

[0097] Briefly, utilizing a commercially available Universal Genome Walker Kit [Clontech], genomic DNAs from monkey tissues or cell lines that are identified as positive for the presence of AAV7 sequence are digested with Dra I, EcoR V, Pvu II and Stu I endonucleases and ligated to Genome Walker Adaptor to generate 4 individual Genome Walker Libraries (GWLs). Using DNAs from GWLs as templates, AAV7 and adjacent genomic sequences will be PCR-amplified by the adaptor primer 1 (API, provided in the kit) and an AAV7 specific primer 1, followed by a nested PCR using the adaptor primer 2 (AP2) and another AAV7 specific primer 2, both of which are internal to the first set of primers. The major PCR products from the nested PCR are cloned and characterized by sequencing analysis.

[0098] In this experiment, the primers covering the 257 bp or other signature fragment of a generic AAV genome are used for PCR amplification of cellular DNAs extracted from Human and NHP derived cell lines to identify and characterize latent AAV sequences. The identified latent AAV genomes are rescued from the positive cell lines using adenovirus helpers of different species and strains.

[0099] To isolate infectious AAV clones from NHP derived cell lines, a desired cell line is obtained from ATCC and screened by PCR to identify the 257 bp amplicon, i.e., signature region of the invention. The 257 bp PCR product is cloned and serotyped by sequencing analysis. For these cell lines containing the AAV7 sequence, the cells are infected with SV-15, a simian adenovirus purchased from ATCC, human Ad5 or transfected with plasmid construct housing the human Ad genes that are responsible for AAV helper functions. At 48 hour post infection or transfection, the cells are harvested and Hirt DNA is prepared for cloning of AAV7 genome following Xiao et al., 1999, J. Virol, 73:3994-4003.

#### Example 5 - Production of AAV Vectors

[0100] A pseudotyping strategy similar to that of Example 3 for AAV1/7 was employed to produce AAV2 vectors packaged with AAV1, AAV5 and AAV8 capsid proteins. Briefly, recombinant AAV genomes equipped with AAV2 ITRs were packaged by triple transfection of 293 cells with cis-plasmid, adenovirus helper plasmid and a chimeric packaging construct where the AAV2 rep gene is fused with cap genes of novel AAV serotypes. To create the chimeric packaging constructs, the Xho I site of p5E18 plasmid at 3169 bp was ablated and the modified plasmid was restricted with Xba I and Xho I in a complete digestion to remove the AAV2 cap gene and replace it with a 2267 bp Spe I/Xho I fragment containing the AAV8 cap gene [Xiao, W., et al., (1999) J Virol 73, 3994-4003]. A similar cloning strategy was used for creation of chimeric packaging plasmids of AAV2/1 and AAV2/5. All recombinant vectors were purified by the standard CsCl<sub>2</sub> sedimentation method except for AAV2/2, which was purified by single step heparin chromatography.

[0101] Genome copy (GC) titers of AAV vectors were determined by TagMan analysis using probes and primers targeting SV40 poly A region as described previously [Gao, G., et al., (2000) Hum Gene Ther 11, 2079-91].

[0102] Vectors were constructed for each serotype for a number of in vitro and in vivo studies. Eight different transgene cassettes were incorporated into the vectors and recombinant virions were produced for each serotype. The recovery of virus, based on genome copies, is summarized in Table 4 below. The yields of vector were high for each serotype with no consistent differences between serotypes. Data presented in the table are average genome copy yields with standard deviation x 10<sup>13</sup> of multiple production lots of 50 plate (150 mm) transfections.

Table 4 Production of Recombinant Vectors

	Table 4. Production of Recombinant Vectors									
	AAV2/1	AAV2/2	AAV2/5	AAV2/7	AAV2/8					
CMV LacZ	7.30 ± 4.33 (n=9)	4.49 ± 2.89 (n=6)	5.19 ± 5.19 (n=8)	3.42 (n=1)	0.87 (n=1)					
CMV EGFP	6.43 ± 2.42 (n=2)	3.39 ± 2.42 (n=2)	5.55 ± 6.49 (n=4)	2.98 ± 2.66 (n=2)	3.74 ± 3.88 (n=2)					
TBG LacZ	4.18 (n=1)	0.23 (n=1)	0.704 ± 0.43 (n=2)	2.16 (n=1)	0.532 (n=1)					
Alb A1AT	4.67 ± 0.75 (n=2)	4.77 (n=1)	4.09 (n=1)	5.04 (n=1)	2.02 (n=1)					
CB A1AT	0.567 (n=1)	0.438 (n=1)	2.82 (n=1)	2.78 (n=1)	0.816 ± 0.679 (n=2)					
TBG rhCG	8.51 ± 6.65 (n=6)	$3.47 \pm 2.09  (n=5)$	$5.26 \pm 3.85  (n=4)$	6.52 ± 3.08 (n=4)	1.83 ± 0.98 (n=5)					
TBG cFIX	1.24 ± 1.29 (n=3)	0.63 ± 0.394 (n=6)	3.74 ± 2.48 (n=7)	4.05 (n=1)	15.8 ± 15.0 (n=5)					

Example 6 - Serologic Analysis of Pseudotyped Vectors

[0103] C57BL/6 mice were injected with vectors of different serotypes of AAVCBA1AT vectors intramuscularly (5 x

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10<sup>11</sup> GC) and serum samples were collected 34 days later. To test neutralizing and cross-neutralizing activity of sera to each serotype of AAV, sera was analyzed in a transduction based neutralizing antibody assay [Gao, G. P., et al., (1996) *J Virol* **70**, 8934-43]. More specifically, the presence of neutralizing antibodies was determined by assessing the ability of serum to inhibit transduction of 84-31 cells by reporter viruses (AAVCMVEGFP) of different serotypes. Specifically, the reporter virus AAVCMVEGFP of each serotype [at multiplicity of infection (MOI) that led to a transduction of 90% of indicator cells] was pre-incubated with heat-inactivated serum from animals that received different serotypes of AAV or from naïve mice. After 1-hour incubation at 37° C, viruses were added to 84-31 cells in 96 well plates for 48 or 72- hour, depending on the virus serotype. Expression of GFP was measured by Fluorolmagin (Molecular Dynamics) and quantified by Image Quant Software. Neutralizing antibody titers were reported as the highest serum dilution that inhibited transduction to less than 50%.

**[0104]** The availability of GFP expressing vectors simplified the development of an assay for neutralizing antibodies that was based on inhibition of transduction in a permissive cell line (i.e., 293 cells stably expressing E4 from Ad5). Sera to selected AAV serotypes were generated by intramuscular injection of the recombinant viruses. Neutralization of AAV transduction by 1:20 and 1:80 dilutions of the antisera was evaluated (See Table 5 below). Antisera to AAV1, AAV2, AAV5 and AAV8 neutralized transduction of the serotype to which the antiserum was generated (AAV5 and AAV8 to a lesser extent than AAV1 and AAV2) but not to the other serotype (i.e., there was no evidence of cross neutralization suggesting that AAV 8 is a truly unique serotype).

Table 5. Serological Analysis of New AAV Serotypes.

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			% Infection on 84-31 cells with AAVCMVEGFP virus:								
		AA	/2/1	AAV	2/2	AA	/2/5	AAV2/7		AAV2/8	
		Serum	Serum dilution: Serum dilution: Serum dilution:		Serum dilution:		Serum dilution:				
Sera:	Immunization Vector	1/20	1/80	1/20	1/80	1/20	1/80	1/20	1/80	1/20	1/80
Group 1	AAV2/1	0	0	100	100	100	100	100	100	100	100
Group 2	AAV2/2	100	100	0	0	100	100	100	100	100	100
Group 3	AAV2/5	100	100	100	100	16.5	16.5	100	100	100	100
Group 4	AAV2/7	100	100	100	100	100	100	61.5	100	100	100
Group 5	AAV2/8	100	100	100	100	100	100	100	100	26.3	60

**[0105]** Human sera from 52 normal subjects were screened for neutralization against selected serotypes. No serum sample was found to neutralize AAV2/7 and AAV2/8 while AAV2/2 and AAV2/1 vectors were neutralized in 20% and 10% of sera, respectively. A fraction of human pooled IgG representing a collection of 60,000 individual samples did not neutralize AAV2/7 and AAV2/8, whereas AAV2/2 and AAV2/1 vectors were neutralized at titers of serum equal to 1/1280 and 1/640, respectively.

Example 7 - In vivo Evaluation of Different Serotypes of AAV Vectors

[0106] In this study, 7 recombinant AAV genomes, AAV2CBhAIAT, AAV2AlbhAIAt, AAV2CMVrhCG, AAV2TBGrhCG, AAV2TBGcFIX, AAV2CMVLacZ and AAV2TBGLacZ were packaged with capsid proteins of different serotypes. In all 7 constructs, minigene cassettes were flanked with AAV2 ITRs. cDNAs of human  $\alpha$ -antitrypsin (AIAT) [Xiao, W., et al., (1999) J Virol 73, 3994-4003]  $\beta$ -subunit of rhesus monkey choriogonadotropic hormone (CG) [Zoltick, P. W. & Wilson, J. M. (2000) *Mol Ther* 2, 657-9] canine factor IX [Wang, L., et al., (1997) *Proc Natl Acad Sci USA* 94, 11563-6] and bacterial  $\beta$ -glactosidase (i.e., Lac Z) genes were used as reporter genes. For liver-directed gene transfer, either mouse albumin gene promoter (Alb) [Xiao, W. (1999), cited above] or human thyroid hormone binding globulin gene promoter (TBG) [Wang (1997), cited above] was used to drive liver specific expression of reporter genes. In muscle-directed gene transfer experiments, either cytomegalovirus early promoter (CMV) or chicken  $\beta$ -actin promoter with CMV enhancer (CB) was employed to direct expression of reporters.

[0107] For muscle-directed gene transfer, vectors were injected into the right tibialis anterior of 4-6 week old NCR nude or C57BL/6 mice (Taconic, Germantown, NY). In liver-directed gene transfer studies, vectors were infused intraportally into 7-9 week old NCR nude or C57BL/6 mice (Taconic, Germantown, NY). Serum samples were collected intraorbitally at different time points after vector administration. Muscle and liver tissues were harvested at different time points for cryosectioning and Xgal histochemical staining from animals that received the lacZ vectors. For the re-administration experiment, C56BL/6 mice initially received AAV2/1, 2/2, 2/5, 2/7 and 2/8CBAIAT vectors intramuscularly and followed for A1AT gene expression for 7 weeks. Animals were then treated with AAV2/8TBGcFIX intraportally and studied for cFIX gene expression.

[0108] ELISA based assays were performed to quantify serum levels of hA1AT, rhCG and cFIX proteins as described previously [Gao, G. P., et al., (1996) *J Virol* 70, 8934-43; Zoltick, P. W. & Wilson, J. M. (2000) *Mol Ther* 2, 657-9; Wang, L., et al., *Proc Natl Acad Sci U S A* 94, 11563-6]. The experiments were completed when animals were sacrificed for harvest of muscle and liver tissues for DNA extraction and quantitative analysis of genome copies of vectors present in target tissues by TaqMan using the same set of primers and probe as in titration of vector preparations [Zhang, Y., et al., (2001) *Mol Ther* 3, 697-707].

**[0109]** The performance of vectors base on the new serotypes were evaluated in murine models of muscle and liver-directed gene transfer and compared to vectors based on the known serotypes AAV1, AAV2 and AAV5. Vectors expressing secreted proteins (alpha-antitrypsin (A1AT) and chorionic gonadotropin (CG)) were used to quantitate relative transduction efficiencies between different serotypes through ELISA analysis of sera. The cellular distribution of transduction within the target organ was evaluated using lacZ expressing vectors and X-gal histochemistry.

[0110] The performance of AAV vectors in skeletal muscle was analyzed following direct injection into the tibialis anterior muscles. Vectors contained the same AAV2 based genome with the immediate early gene of CMV or a CMV enhanced  $\beta$ -actin promoter driving expression of the transgene. Previous studies indicated that immune competent C57BL/6 mice elicit limited humoral responses to the human A1AT protein when expressed from AAV vectors [Xiao, W., et al., (1999) *J Virol* 73, 3994-4003].

[0111] In each strain, AAV2/1 vector produced the highest levels of A I AT and AAV2/2 vector the lowest, with AAV2/7 and AAV2/8 vectors showing intermediate levels of expression. Peak levels of CG at 28 days following injection of nu/nu NCR mice showed the highest levels from AAV2/7 and the lowest from AAV2/2 with AAV2/8 and AAV2/1 in between. Injection of AAV2/1 and AAV2/7 lacZ vectors yielded gene expression at the injection sites in all muscle fibers with substantially fewer lacZ positive fibers observed with AAV2/2 and AAV 2/8 vectors. These data indicate that the efficiency of transduction with AA V2/7 vectors in skeletal muscle is similar to that obtained with AAV2/1, which is the most efficient in skeletal muscle of the previously described serotypes [Xiao, W. (1999), cited above; Chao, H., et al., (2001) *Mol Ther* 4, 217-22; Chao, H., et al., (2000) *Mol Ther* 2, 619-23].

**[0112]** Similar murine models were used to evaluate liver-directed gene transfer. Identical doses of vector based on genome copies were infused into the portal veins of mice that were analyzed subsequently for expression of the transgene. Each vector contained an AAV2 based genome using previously described liver-specific promoters (i.e., albumin or thyroid hormone binding globulin) to drive expression of the transgene. More particularly, CMVCG and TBGCG minigene cassettes were used for muscle and liver-directed gene transfer, respectively. Levels of rhCG were defined as relative units (RUs x 10<sup>3</sup>). The data were from assaying serum samples collected at day 28, post vector administration (4 animals per group). As shown in Table 3, the impact of capsid proteins on the efficiency of transduction of A1AT vectors in nu/nu and C57BL/6 mice and CG vectors in C57BL/6 mice was consistent (See Table 6).

Table 6. Expression of β-unit of Rhesus Monkey Chorionic Gonadotropin (rhCG)

	Vector	Muscle	Liver
•	AAV2/1	4.5 ± 2.1	1.6 ± 1.0
	AAV2	$0.5\pm0.1$	$0.7\pm0.3$
	AAV2/5	ND*	$4.8\pm0.8$
	AAV2/7	$14.2\pm2.4$	$8.2\pm4.3$
	AAV2/8	$4.0\pm0.7$	$76.0\pm22.8$

<sup>\*</sup> Not determined in this experiment.

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[0113] In all cases, AAV2/8 vectors yielded the highest levels of transgene expression that ranged from 16 to 110 greater than what was obtained with AAV2/2 vectors; expression from AAV2/5 and AAV2/7 vectors was intermediate with AAV2/7 higher than AAV2/5. Analysis of X-Gal stained liver sections of animals that received the corresponding lacZ vectors showed a correlation between the number of transduced cells and overall levels of transgene expression. DNAs extracted from livers of C57BL/6 mice who received the A1AT vectors were analyzed for abundance of vector DNA using real time PCR technology.

**[0114]** The amount of vector DNA found in liver 56 days after injection correlated with the levels of transgene expression (See Table 7). For this experiment, a set of probe and primers targeting the SV40 polyA region of the vector genome was used for TaqMan PCR. Values shown are means of three individual animals with standard deviations. The animals were sacrificed at day 56 to harvest liver tissues for DNA extraction. These studies indicate that AAV8 is the most efficient vector for liver-directed gene transfer due to increased numbers of transduced hepatocytes.

Table 7 - Real Time PCR Analysis for Abundance of AAV Vectors in nu/nu Mouse Liver Following Injection of 1x10<sup>11</sup> Genome Copies of Vector.

AAV vectors/Dose		Genome Copies per Cell
	AAV2/1AlbA1AT	$0.6 \pm 0.36$
	AAV2AlbA1AT	$0.003 \pm 0.001$
	AAV2/5AlbA1AT	$0.83\pm0.64$
	AAV2/7AlbA1AT	$2.2 \pm 1.7$
	AAV2/8AlbA1AT	18 ± 11

[0115] The serologic data described above suggest that AAV2/8 vector should not be neutralized *in vivo* following immunization with the other serotypes. C57BL/6 mice received intraportal injections of AAV2/8 vector expressing canine factor IX ( $10^{11}$  genome copies) 56 days after they received intramuscular injections of A1AT vectors of different serotypes. High levels of factor IX expression were obtained 14 days following infusion of AAV2/8 into naïve animals ( $17\pm2~\mu$ g/ml, n=4) which were not significantly different that what was observed in animals immunized with AAV2/1 ( $31\pm23~\mu$ g/ml, n=4), AAV2/2 ( $16~\mu$ g/ml, n=2), and ÅAV2/7( $12~\mu$ g/ml, n=2). This contrasts to what was observed in AAV2/8 immunized animals that were infused with the AAV2/8 factor IX vector in which no detectable factor IX was observed (<  $0.1~\mu$ g/ml, n=4). [0116] Oligonucleotides to conserved regions of the cap gene did amplify sequences from rhesus monkeys that represented unique AAVs. Identical cap signature sequences were found in multiple tissues from rhesus monkeys derived from at least two different colonies. Full-length rep and cap open reading frames were isolated and sequenced from single sources. Only the cap open reading frames of the novel AAVs were necessary to evaluate their potential as vectors because vectors with the AAV7 or AAV8 capsids were generated using the ITRs and rep from AAV2. This also simplified the comparison of different vectors since the actual vector genome is identical between different vector serotypes. In fact, the yields of recombinant vectors generated using this approach did not differ between serotypes.

**[0117]** Vectors based on AAV7 and AAV8 appear to be immunologically distinct (i.e., they are not neutralized by antibodies generated against other serotypes). Furthermore, sera from humans do not neutralize transduction by AAV7 and AAV8 vectors, which is a substantial advantage over the human derived AAVs currently under development for which a significant proportion of the human population has pre-existing immunity that is neutralizing [Chirmule, N., et al., (1999) *Gene Ther* 6, 1574-83].

[0118] The tropism of each new vector is favorable for *in vivo* applications. AAV2/7 vectors appear to transduce skeletal muscle as efficiently as AAV2/1, which is the serotype that confers the highest level of transduction in skeletal muscle of the primate AAVs tested to date [Xiao, W., cited above; Chou (2001), cited above, and Chou (2000), cited above]. Importantly, AAV2/8 provides a substantial advantage over the other serotypes in terms of efficiency of gene transfer to liver that until now has been relatively disappointing in terms of the numbers of hepatocytes stably transduced. AAV2/8 consistently achieved a 10 to 100-fold improvement in gene transfer efficiency as compared to the other vectors. The basis for the improved efficiency of AAV2/8 is unclear, although it presumably is due to uptake via a different receptor that is more active on the basolateral surface of hepatocytes. This improved efficiency will be quite useful in the development of liver-directed gene transfer where the number of transduced cells is critical, such as in urea cycle disorders and familial hypercholesterolemia.

**[0119]** Thus, the present invention provides a novel approach for isolating new AAVs based on PCR retrieval of genomic sequences. The amplified sequences were easily incorporated into vectors and tested in animals. The lack of pre-existing immunity to AAV7 and the favorable tropism of the vectors for muscle indicates that AAV7 is suitable for use as a vector in human gene therapy and other *in vivo* applications. Similarly, the lack of pre-existing immunity to the AAV serotypes of the invention, and their tropisms, renders them useful in delivery of therapeutic molecules and other useful molecules.

#### Example 9 - Tissue Tropism Studies

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[0120] In the design of a high throughput functional screening scheme for novel AAV constructs, a non-tissue specific and highly active promoter, CB promoter (CMV enhanced chicken  $\beta$  actin promoter) was selected to drive an easily detectable and quantifiable reporter gene, human  $\alpha$  anti-trypsin gene. Thus only one vector for each new AAV clone needs to be made for gene transfer studies targeting 3 different tissues, liver, lung and muscle to screen for tissue tropism of a particular AAV construct. The following table summarizes data generated from 4 novel AAV vectors in the tissue tropism studies (AAVCBA1AT), from which a novel AAV capsid clone, 44.2, was found to be a very potent gene transfer vehicle in all 3 tissues with a big lead in the lung tissue particularly. Table 8 reports data obtained (in  $\mu$ g A1AT/mL serum) at day 14 of the study.

Table 8

Vector	Target Tissue					
	Lung	Liver	Muscle			
AAV2/1	ND	ND	45±11			
AAV2/5	0.6±0.2	ND	ND			
AAV2/8	ND	84±30	ND			
AAV2/rh.2 (43.1)	14±7	25±7.4	35±14			
AAV2/rh.10 (44.2)	23±6	53±19	46±11			
AAV2/rh.13 (42.2)	3.5±2	2±0.8	3.5±1.7			
AAV2/rh.21 (42.10)	3.1±2	2±1.4	4.3±2			

A couple of other experiments were then performed to confirm the superior tropism of AAV 44.2 in lung tissue. First, AAV vector carried CC10hA1AT minigene for lung specific expression were pseudotyped with capsids of novel AAVs were given to Immune deficient animals (NCR nude) in equal volume (50  $\mu$ I each of the original preps without dilution) via intratracheal injections as provided in the following table. In Table 9, 50  $\mu$ I of each original prep per mouse, NCR Nude, detection limit  $\geq$ 0.033  $\mu$ g/mI, Day 28

Table 9

		Table 9		
Vector	Total GC in 50 μl vector	μg of A1AT/ml with 50μl vector	μg of A1AT/ml with 1x10 <sup>11</sup> vector	Relative Gene transfer as compared to rh.10 (clone 44.2)
2/1	3x10 <sup>12</sup>	2.6±0.5	0.09±0.02	2.2
2/2	5.5x10 <sup>11</sup>	<0.03	<0.005	<0.1
2/5	3.6x10 <sup>12</sup>	0.65±0.16	0.02±0.004	0.5
2/7	4.2x10 <sup>12</sup>	1±0.53	0.02±0.01	0.5
2/8	7.5x10 <sup>11</sup>	0.9±0.7	0.12±0.09	2.9
2/ch.5 (A.3.1)	9x10 <sup>12</sup>	1±0.7	0.01±0.008	0.24
2/rh.8 (43.25)	4.6x10 <sup>12</sup>	26±21	0.56±0.46	13.7
2/rh.10 (44.2)	2.8x10 <sup>12</sup>	115±38	4.1±1.4	100
2/rh.13 (42.2)	6x10 <sup>12</sup>	7.3±0.8	0.12±0.01	2.9
2/rh.21 (42.10)	2.4x10 <sup>12</sup>	9±0.9	0.38±0.04	9.3
2/rh.22 (42.11)	2.6x10 <sup>12</sup>	6±0.4	0.23±0.02	5.6
2/rh.24 (42.13)	1.1x10 <sup>11</sup>	0.4±0.3	0.4±0.3	1

The vectors were also administered to immune competent animals (C57BL/6) in equal genome copies (1x10<sup>11</sup> GC) as shown in the Table 10. (1x10<sup>11</sup> GC per animal, C57BL/6, day 14, detection limit  $\geq$ 0.033  $\mu$ g/ml)

Table 10

AAV Vector	μg of A1AT/ml with 1x10 <sup>11</sup> vector	Relative Gene transfer as compared to rh.10 (clone 44.2)						
2/1	0.076±0.031	2.6						
2/2	0.1±0.09	3.4						
2/5	0.0840.033	2.9						

Table continued

AAV Vector	μg of A1AT/ml with 1x10 <sup>11</sup> vector	Relative Gene transfer as compared to rh.10 (clone 44.2)
2/7	0.33±0.01	11
2/8	1.92±1.3	2.9
2/ch.5 (A.3.1)	0.048±0.004	1.6
2/rh.8 (43.25)	1.7±0.7	58
2/rh.10 (44.2)	2.93±1.7	100
2/rh.13 (42.2)	0.45±0.15	15
2/rh.21 (42.10)	0.86±0.32	29
2/rh.22 (42.11)	0.38±0.18	13
2/rh.24 (42.13)	0.3±0.19	10

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[0121] The data from both experiments confirmed the superb tropism of clone 44.2 in lung-directed gene transfer.

[0122] Interestingly, performance of clone 44.2 in liver and muscle directed gene transfer was also outstanding, close to that of the best liver transducer, AAV8 and the best muscle transducer AAV1, suggesting that this novel AAV has some intriguing biological significance.

**[0123]** To study serological properties of those novel AAVs, pseudotyped AAVGFP vectors were created for immunization of rabbits and in vitro transduction of 84-31 cells in the presence and absence of antisera against different capsids. The data are summarized below:

Table 11a. Cross-NAB assay in 8431 cells and adenovirus (Adv) coinfection Infection in 8431 cells (coinfected with Adv) with:

00	Serum from rabbit immunized with:	10 <sup>9</sup> GC	10 <sup>9</sup> GC	10 <sup>9</sup> GC	10 <sup>10</sup> GC
30		rh.13	rh.21	rh.22	rh.24
		AAV2/42.2	AAV2/42.10	AAV2/42.1	AAV2/42.13
	AAV2/1	1/20	1/20	1/20	No NAB
35	AAV2/2	1/640	1/1280	1/5120	No NAB
	AAV2/5	No NAB	1/40	1/160	No NAB
	AAV2/7	1/81920	1/81920	1/40960	1/640
40	AAV2/8	1/640	1/640	1/320	1/5120
40	<b>Ch.5</b> AAV2/A3	1/20	1/160	1/640	1/640
	rh.8				
	AAV2/43.25	1/20	1/20	1/20	1/320
45	rh.10				
	AAV2/44.2	No NAB	No NAB	No NAB	1/5120
	<b>rh.13</b> AAV2/42.2	1/5120	1/5120	1/5120	No NAB
50	rh.21				
	AA V2/42.10	1/5120	1/10240	1/5120	1/20
	rh.22				
	AAV2/42.11	1/20480	1/20480	1/40960	No NAB
55	<b>rh.24</b> AAV2/42.13	No NAB	1/20	1/20	1/5120

Table 11b. Cross-NAB assay in 8431 cells and Adv coinfection Infection in 8431 cells (coinfected with Adv) with:

Serum from rabbit immunized with:	10 <sup>9</sup> GC	10 <sup>10</sup> GC	10 <sup>10</sup> GC	10 <sup>9</sup> GC	10 <sup>9</sup> GC
	rh.12	ch.5	rh. 8	rh.10	rh.20
	AAV2/42.1B	AAV2/A3	AAV2/43.25	AAV2/44.2	AAV2/42.8.2
AAV2/1	No NAB	1/20480	No NAB	1/80	ND
AAV2/2	1/20	No NAB	No NAB	No NAB	ND
AAV2/5	No NAB	1/320	No NAB	No NAB	ND
AAV2/7	1/2560	1/640	1/160	1/81920	ND
AAV2/8	1/10240	1/2560	1/2560	1/81920	ND
<b>ch.5</b> AAV2/A3	1/1280	1/10240	ND	1/5120	1/320
<b>rh.8</b> AAV2/43.25	1/1280	ND	1/20400	1/5120	1/2560
<b>rh.10</b> AAV2/44.2	1/5120	ND	ND	1/5120	1/5120
<i>rh.13</i> AAV2/42.2	1/20	ND	ND	No NAB	1/320
<i>rh.21</i> AAV2/42.10	1/20	ND	ND	1/40	1/80
<i>rh.22</i> AAV2/42.1 1	No NAB	ND	ND	ND	No NAB
<i>rh.24</i> AAV2/42.13	1/5120	ND	ND	ND	1/2560

Table 12

	Titer of rabbit s	Titer after Boosting				
	Vector	Titer d21				
ch.5	AAV2/A3	1/10,240	1/40,960			
rh.8	AAV2/43.25	1/20,400	1/163,840			
rh.10	AAV2/44.2	1/10,240	1/527,680			
rh.13	AAV2/42.2	1/5,120	1/20,960			
rh.21	AAV2/42.10	1/20,400	1/81,920			
rh.22	AAV2/42.11	1/40,960	N D			
rh.24	AAV2/42.13	1/5,120	ND			

Table 13 a. Infection in 8431 cells (coinfected with Adv) with GFP

	10 <sup>9</sup> GC/well					
						ch.5
	AAV2/1	AAV2/2	AAV2/5	AAV2/7	AAV2/8	AAV2/A3
	128	>200	95	56	13	1
# GFU/field	83	>200	65	54	11	1

Table 13b. Infection in 8431 cells (coinfected with Adv) with GFP

	109 GC/well	10 <sup>9</sup> GC/well	10 <sup>9</sup> GC/well	109 GC/well	109 GC/well	10 <sup>9</sup> GC/well	109 GC/well
	rh.8	rh.10	rh.13	rh.21	rh.22	rh.24	rh.12
	AAV2/43.25	AAV2/44.2	AAV2/42.2	AAV2/42.10	AAV2/42.11	AAV2/42.13	AAV2/42.1B
	3	13	54	62	10	3	18
# GFU/field	2	12	71	60	14	2	20
			48	47	16	3	12

#### Example 10 - Mouse Model of Familial Hypercholesterolemia

**[0124]** The following experiment demonstrates that the AAV2/7 construct of the invention delivers the LDL receptor and express LDL receptor in an amount sufficient to reduce the levels of plasma cholesterol and triglycerides in animal models of familial hypercholesterolemia.

#### A. Vector Construction

[0125] AAV vectors packaged with AAV7 or AAV8 capsid proteins were constructed using a pseudotyping strategy [Hildinger M, et al., J. Virol 2001; 75:6199-6203]. Recombinant AAV genomes with AAV2 inverted terminal repeats (ITR) were packaged by triple transfection of 293 cells with the *cis*-plasmid, the adenovirus helper plasmid and a chimeric packaging construct, a fusion of the capsids of the novel AAV serotypes with the rep gene of AAV2. The chimeric packaging plasmid was constructed as previously described [Hildinger et al, cited above]. The recombinant vectors were purified by the standard CsCl<sub>2</sub> sedimentation method. To determine the yield TaqMan (Applied Biosystems) analysis was performed using probes and primers targeting the SV40 poly(A) region of the vectors [Gao GP, et al., Hum Gene Ther. 2000 Oct 10;11(15):2079-91]. The resulting vectors express the transgene under the control of the human thyroid hormone binding globulin gene promoter (TBG).

#### B. Animals

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[0126] LDL receptor deficient mice on the C57Bl/6 background were purchased from the Jackson Laboratory (Bar Harbor, ME, USA) and maintained as a breeding colony. Mice were given unrestricted access to water and obtained a high fat Western Diet (high % cholesterol) starting three weeks prior vector injection. At day -7 as well at day 0, blood was obtained via retroorbital bleeds and the lipid profile evaluated. The mice were randomly divided into seven groups. The vector was injected via an intraportal injection as previously described ([Chen SJ et al., Mol Therapy 2000; 2(3), 256-261]. Briefly, the mice were anaesthetized with ketamine and xylazine. A laparotomy was performed and the portal vein exposed. Using a 30g needle the appropriate dose of vector diluted in 100ul PBS was directly injected into the portal vein. Pressure was applied to the injection site to ensure a stop of the bleeding. The skin wound was closed and draped and the mice carefully monitored for the following day. Weekly bleeds were performed starting at day 14 after liver directed gene transfer to measure blood lipids. Two animals of each group were sacrificed at the time points week 6 and week 12 after vector injection to examine atherosclerotic plaque size as well as receptor expression. The remaining mice were sacrificed at week 20 for plaque measurement and determination oftransgene expression.

Table 14

	Table 14		
	Vector	dose	n
Group 1	AAV2/7-TBG-hLDLr	1x 10 <sup>12</sup> gc	12
Group 2	AAV2/7-TBG-hLDLr	3x 10 <sup>11</sup> gc	12
Group 3	AAV2/7-TBG-hLDLr	1x 10 <sup>11</sup> gc	12
Group 4	AAV2/8-TBG-hLDLr	1x 10 <sup>12</sup> gc	12
Group 5	AAV2/8-TBG-hLDLr	3x 10 <sup>11</sup> gc	12
Group 6	AAV2/8-TBG-hLDLr	1x 10 <sup>11</sup> gc	12
Group 7	AAV2/7-TBG-LacZ	1x 10 <sup>11</sup> gc	16

#### C. Serum lipoprotein and liver function analysis

**[0127]** Blood samples were obtained from the retroorbital plexus after a 6 hour fasting period. The serum was separated from the plasma by centrifugation. The amount of plasma lipoproteins and liver transaminases in the serum were detected using an automatized clinical chemistry analyzer (ACE, Schiapparelli Biosystems, Alpha Wassermann)

#### D. Detection of transgene expression

**[0128]** LDL receptor expression was evaluated by immuno-fluorescence staining and Western blotting. For Western Blot frozen liver tissue was homogenized with lysis buffer (20 mM Tris, pH7.4, 130mM NaCl, 1% Triton X 100, proteinase inhibitor (complete, EDTA-free, Roche, Mannheim, Germany). Protein concentration was determined using the Micro

BCA Protein Assay Reagent Kit (Pierce, Rockford, IL). 40 μg of protein was resolved on 4- 15% Tris-HCl Ready Gels (Biorad, Hercules, CA) and transferred to a nitrocellulose membrane (Invitrogen,). To generate Anti-hLDL receptor antibodies a rabbit was injected intravenously with an AdhLDLr prep (1x10<sup>13</sup> GC). Four weeks later the rabbit serum was obtained and used for Western Blot. A 1:100 dilution of the serum was used as a primary antibody followed by a HRP-conjugated anti-rabbit IgG and ECL chemiluminescent detection (ECL Western Blot Detection Kit, Amersham, Arlington Heights, IL).

#### E. Immunocytochemistry

[0129] For determination of LDL receptor expression in frozen liver sections immunohistochemistry analyses were performed. 10um cryostat sections were either fixed in acetone for 5 minutes, or unfixed. Blocking was obtained *via a* 1 hour incubation period with 10% of goat serum. Sections were then incubated for one hour with the primary antibody at room temperature. A rabbit polyclonal antibody anti-human LDL (Biomedical Technologies Inc., Stoughton, MA) was used diluted accordingly to the instructions of the manufacturer. The sections were washed with PBS, and incubated with 1:100 diluted fluorescein goat anti-rabbit IgG (Sigma, St Louis, MO). Specimens were finally examined under fluorescence microscope Nikon Microphot-FXA. In all cases, each incubation was followed by extensive washing with PBS. Negative controls consisted of preincubation with PBS, omission of the primary antibody, and substitution of the primary antibody by an isotype-matched non-immune control antibody. The three types of controls mentioned above were performed for each experiment on the same day.

#### F. Gene transfer efficiency

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**[0130]** Liver tissue was obtained after sacrificing the mice at the designated time points. The tissue was shock frozen in liquid nitrogen and stored at -80 °C until further processing. DNA was extracted from the liver tissue using a QIAamp DNA Mini Kit (QIAGEN GmbH, Germany) according to the manufacturers protocol. Genome copies of AAV vectors in the liver tissue were evaluated using Taqman analysis using probes and primers against the SV40 poly(A) tail as described above.

#### G. Atherosclerotic plaque measurement

[0131] For the quantification of the atherosclerotic plaques in the mouse aorta the mice were anaesthetized (10% ketamine and xylazine, ip), the chest opened and the arterial system perfused with ice-cold phosphate buffered saline through the left ventricle. The aorta was then carefully harvested, slit down along the ventral midline from the aortic arch down to the femoral arteries and fixed in formalin. The lipid-rich atherosclerotic plaques were stained with Sudan IV (Sigma, Germany) and the aorta pinned out flat on a black wax surface. The image was captured with a Sony DXC-960 MD color video camera. The area of the plaque as well as of the complete aortic surface was determined using Phase 3 Imaging Systems (Media Cybernetics).

## H. Clearance of I125 LDL

[0132] Two animals per experimental group were tested. A bolus of  $I^{125}$ -labeled LDL (generously provided by Dan Rader, U Penn) was infused slowly through the tail vein over a period of 30 sec (1,000,000 counts of  $I^{125}$ )-LDL diluted in  $100\mu$ I sterile PBS/ animal). At time points 3min, 30 min, 1.5hr, 3hr, 6hr after injection a blood sample was obtained *via* the retro-orbital plexus. The plasma was separated off from the whole blood and  $10\mu$ I plasma counted in the gamma counter. Finally the fractional catabolic rate was calculated from the lipoprotein clearance data.

#### I. Evaluation of Liver Lipid accumulation

**[0133]** Oil Red Staining of frozen liver sections was performed to determine lipid accumulation. The frozen liver sections were briefly rinsed in distilled water followed by a 2 minute incubation in absolute propylene glycol. The sections were then stained in oil red solution (0.5% in propylene glycol) for 16 hours followed by counterstaining with Mayer's hematoxylin solution for 30 seconds and mounting in warmed glycerin jelly solution.

**[0134]** For quantification of the liver cholesterol and triglyceride content liver sections were homogenized and incubated in chloroform/methanol (2:1) overnight. After adding of 0.05% H $_2$ SO $_4$  and centrifugation for 10 minutes, the lower layer of each sample was collected, divided in two aliquots and dried under nitrogen. For the cholesterol measurement the dried lipids of the first aliquot were dissolved in 1% Triton X-100 in chloroform. Once dissolved, the solution was dried under nitrogen. After dissolving the lipids in ddH $_2$ 0 and incubation for 30 minutes at 37 °C the total cholesterol concentration was measured using a Total Cholesterol Kit (Wako Diagnostics). For the second aliquot the dried lipids were dissolved

in alcoholic KOH and incubated at 60°C for 30 minutes. Then 1 M MgCl2 was added, followed by incubation on ice for 10 minutes and centrifugation at 14,000 rpm for 30 minutes. The supernatant was finally evaluated for triglycerides (Wako Diagnostics).

[0135] All of the vectors pseudotyped in an AAV2/8 or AAV2/7 capsid lowered total cholesterol, LDL and triglycerides as compared to the control. These test vectors also corrected phenotype of hypercholesterolemia in a dose-dependent manner. A reduction in plaque area for the AAV2/8 and AAV2/7 mice was observed in treated mice at the first test (2 months), and the effect was observed to persist over the length of the experiment (6 months).

Example 10 - Functional Factor IX Expression and Correction of Hemophilia

A. Knock-Out Mice

**[0136]** Functional canine factor IX (FIX) expression was assessed in hemophilia B mice. Vectors with capsids of AAV1, AAV2, AAV5, AAV7 or AAV8 were constructed to deliver AAV2 5' ITR - liver-specific promoter [LSP] - canine FIX - woodchuck hepatitis post-regulatory element (WPRE) - AAV2 3' ITR. The vectors were constructed as described in Wang et al, 2000, *Molecular Therapy* 2: 154-158), using the appropriate capsids.

[0137] Knock-out mice were generated as described in Wang et al, 1997. *Proc. Natl. Acad. Sci. USA* 94: 11563-11566. This model closely mimic the phenotypes of hemophilia B in human.

**[0138]** Vectors of different serotypes (AAV1, AAV2, AAV5, AAV7 and AAV8) were delivered as a single intraportal injection into the liver of adult hemophiliac C57Bl/6 mice in a dose of 1x10<sup>11</sup> GC/mouse for the five different serotypes and one group received an AAV8 vector at a lower dose, 1x10<sup>10</sup> GC/mouse. Control group was injected with 1 x 10<sup>11</sup> GC of AAV2/8 TBG LacZ3. Each group contains 5-10 male and female mice. Mice were bled bi-weekly after vector administration.

1. ELISA

[0139] The canine FIX concentration in the mouse plasma was determined by an ELISA assay specific for canine factor IX, performed essentially as described by Axelrod et al, 1990, *Proc.Natl.Acad.Sci. USA, 87:5173-5177* with modifications. Sheep anti-canine factor IX (Enzyme Research Laboratories) was used as primary antibody and rabbit anti-canine factor IX ((Enzyme Research Laboratories) was used as secondary antibody. Beginning at two weeks following injection, increased plasma levels of cFIX were detected for all test vectors. The increased levels were sustained at therapeutic levels throughout the length of the experiment, i.e., to 12 weeks. Therapeutic levels are considered to be 5% of normal levels, i.e., at about 250 ng/mL.

**[0140]** The highest levels of expression were observed for the AAV2/8 (at 10<sup>11</sup>) and AAV2/7 constructs, with sustained superphysiology levels cFIX levels (ten-fold higher than the normal level). Expression levels for AAV2/8 (10<sup>11</sup>) were approximately 10 fold higher than those observed for AAV2/2 and AAV2/8 (10<sup>10</sup>). The lowest expression levels, although still above the therapeutic range, were observed for AAV2/5.

2. In Vitro Activated Partial Thromboplastin time (aPTT) Assay

**[0141]** Functional factor IX activity in plasma of the FIX knock-out mice was determined by an *in vitro* activated partial thromboplastin time (aPTT) assay-Mouse blood samples were collected from the retro-orbital plexus into 1/10 volume of citrate buffer. The aPTT assay was performed as described by Wang et al, 1997, *Proc. Natl. Acad. Sci. USA* **94**: 11563-11566.

[0142] Clotting times by aPTT on plasma samples of all vector injected mice were within the normal range (approximately 60 sec) when measured at two weeks post-injection, and sustained clotting times in the normal or shorter than normal range throughout the study period (12 weeks).

**[0143]** Lowest sustained clotting times were observed in the animals receiving AAV2/8 (10<sup>11</sup>) and AAV2/7. By week 12, AAV2/2 also induced clotting times similar to those for AAV2/8 and AAV2/7. However, this lowered clotting time was not observed for AAV2/2 until week 12, whereas lowered clotting times (in the 25 - 40 sec range) were observed for AAV2/8 and AAV2/7 beginning at week two.

[0144] Immuno-histochemistry staining on the liver tissues harvested from some of the treated mice is currently being performed. About 70-80% of hepatocytes are stained positive for canine FIX in the mouse injected with AAV2/8.cFIX vector.

B. Hemophilia B Dogs

[0145] Dogs that have a point mutation in the catalytic domain of the F.IX gene, which, based on modeling studies,

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appears to render the protein unstable, suffer from hemophilia B [Evans et al, 1989, Proc. Natl. Acad. Sci. USA, 86:10095-10099). A colony of such dogs has been maintained for more than two decades at the University of North Carolina, Chapel Hill. The homeostatic parameters of these dogs are well described and include the absence of plasma F.IX antigen, whole blood clotting times in excess of 60 minutes, whereas normal dogs are 6-8 minutes, and prolonged activated partial thromboplastin time of 50-80 seconds, whereas normal dogs are 13-28 seconds. These dogs experience recurrent spontaneous hemorrhages. Typically, significant bleeding episodes are successfully managed by the single intravenous infusion of 10 ml/kg of normal canine plasma; occasionally, repeat infusions are required to control bleeding. [0146] Four dogs are injected intraportally with AAV.cFIX according to the schedule below. A first dog receives a single injection with AAV2/2.cFIX at a dose of 3.7x10<sup>11</sup> genome copies (GC)/kg. A second dog receives a first injection of AAV2/2.cFIX (2.8x10<sup>11</sup> GC/kg), followed by a second injection with AAV2/7.cFIX (2.3x10<sup>13</sup> GC/kg) at day 1180. A third dog receives a single injection with AAV2/2.cFIX at a dose of 4.6x10<sup>12</sup> GC/kg. The fourth dog receives an injection with AAV2/2.cFIX (2.8x10<sup>12</sup> GC/kg) and an injection at day 99.5 with AAV2/7.cFIX (5x10<sup>12</sup> GC/kg).

[0147] The abdomen of hemophilia dogs are aseptically and surgically opened under general anesthesia and a single infusion of vector is administered into the portal vein. The animals are protected from hemorrhage in the peri-operative period by intravenous administration of normal canine plasma. The dog is sedated, intubated to induce general anesthesia, and the abdomen shaved and prepped. After the abdomen is opened, the spleen is moved into the operative field. The splenic vein is located and a suture is loosely placed proximal to a small distal incision in the vein. A needle is rapidly inserted into the vein, then the suture loosened and a 5 F cannula is threaded to an intravenous location near the portal vein bifurcation. After hemostasis is secured and the catheter balloon inflated, approximately 5.0 ml of vector diluted in PBS is infused into the portal vein over a 5 minute interval. The vector infusion is followed by a 5.0 ml infusion of saline. The balloon is then deflated, the callula removed and venous hemostasis is secured. The spleen is then replaced, bleeding vessels are cauterized and the operative wound is closed. The animal is extubated having tolerated the surgical procedure well. Blood samples are analyzed as described. [Wang et al, 2000, *Molecular Therapy* 2: 154-158]

[0148] Results showing correction or partial correction are anticipated for AAV2/7.

SEQUENCE LISTING

#### [0149]

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210

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<213> adeno-associated virus serotype 2

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<213> new AAV serotype, clone 42.4

<400> 33

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<213> new AAV serotype, clone 223.2

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<211> 1933

<212> DNA

<213> new AAV serotype, clone 223.4

<400> 50

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<211> 1933 <212> DNA

<213> new AAV serotype, clone 223.6

<400> 52

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<210> 54

<211> 3123

<212> DNA <213> new AAV serotype, clone A3.4

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<400> 54

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<212> DNA <213> new AAV serotype, clone A3.5

<400> 55

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<211> 3122

<212> DNA

<213> new AAV serotype, clone A3.7

<400> 56

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<211> 3129

<212> DNA

<213> new AAV serotype, clone 44.2

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<212> PRT

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25	Asp	Asn	Leu	Gln 580	Gln	Gln	Asn	Ala	Ala 585	Pro	Ile	Val	Gly	Ala 590	Val	Asn
30	Ser	Gln	Gly 595	Ala	Leu	Pro	Gly	Met 600	Ala	Trp	Gln	Asn	Arg 605	Asp	Val	Tyr
	Leu	Gln 610	Gly	Pro	Ile	Trp	Ala 615	ГÀа	Ile	Pro	His	Thr 620	Asp	Gly	Asn	Phe
35	His 625	Pro	Ser	Pro	Leu	Met 630	Gly	Gly	Phe	Gly	Leu 635	Lys	His	Pro	Pro	Pro 640
40	Gln	Ile	Leu	Ile	Lys 645	Asn	Thr	Pro	Val	Pro 650	Ala	Asp	Pro	Pro	Thr 655	Thr
	Phe	Ser	Gln	Ala 660	Lys	Leu	Ala	Ser	Phe 665	Ile	Thr	Gln	Tyr	Ser 670	Thr	Gly
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50	Arg	Trp 690	Asn	Pro	Glu	Ile	Gln 695	Tyr	Thr	Ser	Asn	Туг 700	Tyr	Lys	Ser	Thr
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20	Lys	Ala	Asn 35	Gln	Gln	Lys	Gln	Asp 40	Asp	Gly	Arg	Gly	Leu 45	Val	Leu	Pro
25	Gly	Tyr 50	Lys	Tyr	Leu	Gly	Pro 55	Phe	Asn	Gly	Leu	Asp 60	Lys	Gly	Glu	Pro
	<b>Val</b> 65	Asn	Glu	Ala	qeA	Ala 70	Ala	Ala	Leu	Glu	His 75	Asp	Lys	Ala	Tyr	Asp 80
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35	_	Leu		100					105					110		
		Gly	115					120					125			
40		130					135					140				
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		Pro			165					170					175	
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	Asp	210	Asn	Glu	Gly	Ala	Asp 215	Gly	Val	Gly	Ser	ser 220	Ser	Gly	Asn	Trp

	His 225	Суз	qeA	Ser	Thr	Trp 230	Leu	Gly	Asp	Arg	Val 235	Ile	Thr	Thr	Ser	Thr 240
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15	Tyr	Ser	Thr 275	Pro	Trp	Gly	Туг	Phe 280	Asp	Phe	Asn	Arg	Phe 285	His	Суз	His
	Phe	Ser 290	Pro	Arg	Asp	Trp	Gln 295	Arg	Leu	Ile	Asn	Asn 300	Asn	Trp	вју	Phe
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	Val	Thr	Gln	Asn	Glu 325	Gly	Thr	Lys	Thr	Ile 330	Ala	Asn	Asn	Leu	Thr 335	Ser
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	Pro	Phe	His	Ser 420	Ser	Tyr	Ala	His	Ser 425	Gln	Ser	Leu	Asp	Arg 430	Leu	Met
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	Pro 465	Asn	Asn	Met	Ser	Ala 470	Gln	Ala	Lys	Asn	Trp 475	Leu	Pro	Gly	Pro	Cys 480

	Tyr	Arg	Gln	Gln	Arg 485	Val	Ser	Thr	Thr	Val 490	Ser	Gln	Asn	Asn	Asn 495	Ser
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15	Glu	Arg 530		Phe	Pro	Ser	Ser S35	Gly	Val	Leu	Met	Phe 540	GJĄ	Lys	Gln	Gl <b>y</b>
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23	Val	Asn	ser 595	Gln	GJÀ	Ala	Leu	Pro 600	Gly	Met	Val	Trp	Gln 605	Asn	Arg	Ąsp
30	Val	Tyr 610	Leu	Gln	Gly	Pro	Ile 615	Trp	Ala	Lys	Ile	Pro 620	His	Thr	Asp	GĮ <b>À</b>
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35	Pro	Pro	Gln	Ile	Leu 645	Ile	Lys	Asn	Thr	Pro 650	Val	Pro	Ala	Asp	Pro 655	Pro
40	Thr	Thr	Phe	Ser 660	Gln	Ala	Lys	Leu	<b>Ala</b> 665	Ser	Phe	Ile	Thr	Gln 670	Tyr	Ser
	Thr	вĵу	Gln 675	Val	Ser	Val	Glu	Ile 680	Glu	Trp	Glu	Leu	Gln 685	Lys	Glu	Asn
45	Ser	Lys 690	Arg	Trp	Asn	Pro	Glu 695	Ile	Gln	Tyr	Thr	Ser 700	Asn	Tyr	Tyr	Lys
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35		Pro 145	Val	Glu	Pro	Ser	Pro 150	Gln	Arg	Ser	Pro	Asp 155	Ser	Ser	Thr	Gly	Ile 160
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		Gly	Ala 210	Pro	Met	Ala	Asp	Asn 215	Asn	Glu	Gly	Ala	Asp 220	Gly	Val	Gly	Ser
50		Ser 225	Ser	Gly	Asn	Trp	His 230	Cys	Ąsp	Ser		Trp 235	Leu	Gly	qeA	Arg	Val 240
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10	Arg	Phe 290		к Суз	His	Phe	Ser 295		Arg	qeA ı	Trp	Glr 300		Leu	lle	: Asr
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	Ser	Arg 450	Thr	Gln	Ser	Thr	Gly 455	Gly	Thr	Ala	Gly	Thr 460	Gln	Gln	Leu	Leu
45	Phe 465	Ser	Gln	Ala	Gly	Pro 470	Asn	Asn	Met	Ser	Ala 475	Gln	Ala	Lys	Asn	Trp 480
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	Gln	Asn	Asn	Asn 500	Ser	Asn	Phe	Ala	Trp 505	Thr	Gly	Ala	Thr	Lys 510	Tyr	His

	L	eu Ası	o Gly 515		Asp	Ser	Leu	Val 520		Pro	Gly	Val	Ala 525		Ala	Thr
5	H:	is Lys 530		Asp	Glu	Glu	Arg 535		Phe	Pro	Ser	Ser 540		Val	Leu	Met
10		ne Gly 15	, Lys	Gln	Gly	Ala 550	Gly	Lys	Asp	Asn	Val 555		Tyr	Ser	Ser	Val 560
	Me	et Lev	Thr	Ser	Glu 565		Glu	Ile	Lys	Thr 570	Thr	Asn	Pro	Val	Ala 575	Thr
15	G)	u Glr	Tyr	Gly 580	Val	Val	Ala	Asp	Asn 585	Leu	Gln	Gln	Gln	Asn 590	Ala	Ala
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	Tr	p Gln 610		Arg	Asp	Val	Tyr 615	Leu	Gln	Gly	Pro	Ile 620	Trp	Ala	Lys	Ile
25	Pr 62	o His 5	Thr	Asp	Gly	Asn 630	Phe	His	Pro	Ser	Pro 635	Leu	Met	Gly	Gly	Phe 640
30	Gl	y Leu	Lys	His	Pro 645	Pro	Pro	Gln	Ile	Leu 650	Ile	Lys	Asn	Thr	Pro 655	Val
	Pr	o Ala	Asp	Pro 660	Pro	Thr	Thr	Phe	<b>Ser</b> 665	Gln	Ala	Lys	Leu	Ala 670	Ser	Phe
35	11	e Thr	Gln 675	Tyr	Ser	Thr	Gly	Gln 680	Val	Ser	Val	Glu	Ile 685	Glu	Trp	Glu
40	Le	u Gln 690	Lys	Glu	Asn	Ser	Lys 695	Arg	Trp	Asn	Pro	Glu 700	Ile	Gln	Tyr	Thr
	Se 70	r Asn 5	Tyr	Tyr	Lys	Ser 710	Thr	Asn	Val	Asp	Phe 715	Ala	Val	Asn	Thr	Glu 720
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	Pro 145	Val	Glu	Pro	Ser	Pro 150	Gln	Arg	Ser	Pro	<b>Asp</b> 155	Ser	Ser	Thr	Gly	11e 160
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45	Gly	Ala 210	Pro	Met	Ala	qeA	Asn 215	Asn	Glu	Gly	Ala	Asp 220	Gly	Val	Gly	Ser
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10	Arg	Phe 290	His	Cys	His	Phe	Ser 295		Arg	Asp	Trp	Gln 300		Leu	Ile	Asn
	Asn 305		Trp	Gly	Phe	Arg 310		Lys	Arg	Leu	Asn 315		Lys	Leu	Phe	Asn 320
15	Ile	Gln	Val	Lys	Glu 325		Thr	Gln	Asn	Glu 330	Gly	Thr	Lуз	Thr	Ile 335	
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25	Pro	Ala 370	Asp	Val	Phe	Met	Ile 375	Pro	Gln	Tyr	Gly	туг 380	Leu	Thr	Leu	Asn
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	Phe	Pro	Ser	Gln	Met 405	Leu	Arg	Thr	Gly	Asn 410	Asn	Phe	Glu	Phe	Ser 415	Tyr
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	Ser	Arg 450	Thr	Gln	Ser	Thr	Gly 455	Gly	Thr	Gln	Gly	Thr 460	Gln	Gln	Leu	Leu
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50	Leu	Pro	Gly	Pro	Cys 485	Tyr	Arg	Gln	Gln	Arg 490	Val	Ser	Thr	Thr	Leu 495	Ser
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		Pro	Ala	Asp	Pro 660	Pro	Thr	Thr	Phe	Ser 665	Gln	Ala	Lys	Leu	Ala 670	Ser	Phe
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	Asp	Ala	Glu	Phe 100	Gln	Glu	Arg	Leu	Gln 105	Glu	Asp	Thr	Ser	Phe 110	Gly	Gly
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	Ser	Arg 450	Thr	Gln	Ser	Thr	Gly 455	вја	Thr	Gln	Gly	Thr 460	Gln	Gln	Leu	Leu
45	Phe 465	Ser	Gln	Ala	GJĀ	Pro 470	Ala	Asn	Met	Ser	Ala 475	Gln	Ala	Lys	Asn	Trp 480
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		Met	Leu	Thr	Ser	Glu 565	Glu	Glu	Ile	Lуз	Thr 570	Thr	Asn	Pro	Val	Ala 575	Thr
15		Glu	Gln	Tyr	Gly 580	Val	Val	Ala	Asp	<b>Asn</b> <b>5</b> 85	Leu	Gln	Gln	Thr	Asn 590	Gly	Ala
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25		Pro 625	His	Thr	Asp	Gly	Asn 630	Phe	His	Pro	Ser	Pro 635	Leu	Met	Gly	Gly	Phe 640
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<i>35</i>		Pro	Ala	Asp	Pro 660	Pro	Thr	Thr	Phe	Ser 665	Gln	Ala	Lys	Leu	Ala 670	Ser	Phe
33		Ile	Thr	Gln 675	Tyr	Ser	Thr	Gly	Gln 680	Val	Ser	Val	Glu	Ile 685	Glu	Trp	Glu
40		Leu	Gln 690	Lys	Glu	Asn	ser	Lys 695	Arg	Trp	Asn	Pro	Glu 700	Ile	Gln	Туr	Thr
45		Ser 705	Asn	Tyr	Tyr	Lys	ser 710	Thr	Asn	Val	Asp	Phe 715	Ala	Val	Asn	Thr	Glu 720
43		Gly	Thr	Tyr	Ser	Glu 725	Pro	Arg	Pro	Ile	Gl <b>y</b> 730	Thr	Arg	Tyr	Leu	Thr 735	Arg
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10	Lys	Ala	Asn 35	Gln	Gln	Lys	Gln	Asp 40	Asp	Gly	Arg	Gly	Leu 45	Val	Leu	Pro
	GIÀ	Tyr 50	Lys	Tyr	Leu	Gly	Pro 55	Phe	Asn	Gly	Leu	Asp 60	Lys	Gly	Glu	Pro
15	Val 65	Asn	Ala	Ala	Asp	Ala 70	Ala	Ala	Leu	Glu	His 75	qeA	Lys	Ala	Tyr	qeA 08
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	Asp	Ala	Glu	Phe 100	Gln	Glu	Arg	Leu	Gln 105	Glu	Asp	Thr	Ser	Phe 110	Gly	Gly
25	Asn	Leu	Gly 115	Arg	Ala	Val	Phe	Gln 120	Ala	Lys	Lys	Arg	Val 125	Leu	Glu	Pro
30	Leu	Gly 130	Leu	Val	G1u	Glu	Gly 135	Ala	Lys	Thr	Ala	Pro 140	GJÀ	Lуз	Lуз	Arg
	Pro 145	Val	Glu	Pro	Ser	Pro 150	Gln	Arg	Ser	Pro	Asp 155	Ser	Ser	Thr	Gly	Ile 160
35	Gly	Lys	Lys	Gly	Gln 165	Gln	Pro	Ala	Arg	Lys 170	Arg	Leu	Asn	Phe	Gly 175	Gln
40	Thr	Gly	Asp	Ser 180	Glu	Ser	Val	Pro	Азр 185	Pro	Gln	Pro	Leu	Gly 190	Glu	Pro
	Pro	Ala	Ala 195	Pro	Ser	Gly	Val	Gly 200	Pro	Asn	Thr	Met	Ala 205	Ala	Glў	Gly
45	Gly	Ala 210	Pro	Met	Ala	Asp	Asn 215	Asn	Glu	СЈĀ	Ala	Asp 220	Gly	Val	GÌУ	Ser
50	Ser 225	Ser	Gly	Asn	Trp	His 230	Суз	Asp	Ser	Thr	Trp 235	Leu	GЉ	Asp	Arg	Val 240

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15	Arg Ph 29		ys His	Phe	Ser 295	Pro	Arg	Asp	Trp	Gln 300	Arg	Leu	Ile	Asn
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40	Thr Ph		Asp Val 420	Pro	Phe	His	Ser 425	Ser	Tyr	Ala	His	ser 430	Gln	Ser
45	Leu As	p Arg I 435	Leu Met	Asn	Pro	Leu 440	Ile	Asp	Gln	туг	Leu 445	Tyr	Tyr	Leu
	Ser Ar 45		Sln Thr	Thr	Gly 455	ely	Thr	Ala	Asn	Thr 460	Gln	Thr	Leu	Gly
50	Phe Se 465	r Gln (	Gly Gly	Pro 470	Asn	Thr	Met	Ala	Asn 475	Gln	Ala	Lys	Asn	Trp 480
55	Leu Pr	o Gly 1	Pro Cys 485		Arg	Gln	Gln	Arg 490	Val	Ser	Thr	Thr	Thr 495	Gly

	Gln	Asn	Asn	Asn 500	Ser	Asn	Phe	Ala	Trp 505	Thr	Ala	Gly	Thr	Lys 510	Tyr	His
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10	His	Lys 530	Asp	Asp	Glu	Glu	Arg 535	Phe	Phe	Pro	Ser	Asn 540	Gly	Ile	Leu	Ile
.e	Phe 545	Gly	Lys	Gln	Asn	Ala 550	Ala	Arg	Asp	Asn	Ala 555	Asp	Tyr	Ser	Asp	<b>Val</b> 560
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20	Glu	Glu	Tyr	Gly 580	Ile	Val	Ala	Ąsp	Asn 585	Leu	Gln	Gln	Gln	Asn 590	Thr	Ala
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25	Trp	Gln 610	Asn	Arg	Asp	Val	Tyr 615	Leu	Gln	Gly	Pro	11e 620	Trp	Ala	Lys	Ile
30	Pro 625	His	Thr	Asp	Gly	Asn 630	Phe	His	Pro	Ser	Pro 635	Leu	Met	Gly	ely	Phe 640
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45	Leu	Gln 690	Lys	Glu	Asn	Ser	Lys 695	Arg	Trp	Asn	Pro	Glu 700	Ile	Gln	Tyr	Thr
45	Ser 705	Asn	Tyr	Tyr	Lys	Ser 710	Thr	Ser	Val	Asp	Phe 715	Ala	Val	Asn	Thr	Glu 720
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<213> capsid protein of AAV serotype, clone 43.21

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	Pro 145	Val	Glu	Gln	Ser	Pro 150	Gln	Glu	Pro	Asp	Ser 155	Ser	Ser	Gly	Ile	Gly 160
40	ГÀЗ	Thr	Gly	Gln	Gln 165	Pro	Ala	Lys	Lys	Arg 170	Leu	Asn	Phe	Gly	Gln 175	Thr
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15	Phe	His 290	Суз	His	Phe	Ser	Pro 295	Arg	Asp	Trp	Gln	Arg 300	Leu	Ile	Asn	Asn
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	Ala	Asp 370	Val	Phe	Met	Val	Pro 375	Gln	Tyr	Gly	Tyr	Leu 380	Thr	Leu	Asn	Asn
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10	Gly	Arg	Asp 515	Ser	Leu	Met	Asn	Pro 520	Gly	Val	Ala	Met	Ala 525		His	Lys
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15	Lys 545		Gly	Ala	Gly	Asn 550	Asp	Gly	Val	Asp	Tyr 555	Ser	Gln	Val	Leu	Ile 560
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	Thr 625	Asp	Gly	Asn	Phe	His 630	Pro	Ser	Pro	Leu	Met 635	Gly	Gly	Phe	Gly	Leu 640
35	Lys	His	Pro	Pro	Pro 645	Gln	Ile	Leu	Ile	Lys 650	Asn	Thr	Pro	Val	Pro 655	Ala
40	Asp	Pro	Pro	Leu 660	Thr	Phe	Asn	Gln	Ala 665	Lys	Leu	Asn	Ser	Phe 670	Ile	Thr
_	Gln	Tyr	Ser 675	Thr	Gly	Gln	Val	Ser 680	Val	Glu	Ile	<b>G</b> lu	Trp 685	Glu	Leu	Gln
45	Lys	Glu 690	Asn	Ser	Lys	Arg	Trp 695	Asn	Pro	Glu	Ile	Gln 700	Tyr	Thr	Ser	Asn
50	Tyr 705	Tyr	Lys	Ser		Asn 710	Val .	Asp	Phe	Ala	<b>Val</b> 715	Asn	Thr	Glu	Gly	Val 720
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<213> capsid protein of AAV serotype, clone 43.25

<400> 97

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15	Гуз	Ala	Asn 35	Gln	Gln	Lys	Gln	Asp 40	qeA o	Gly	Arg	Gly	Leu 45	Val	Leu	Pro
	Gly	Tyr 50	Lys	Туг	Leu	Gly	Pro 55	Phe	Asn	Gly	ren	Asp 60	Lys	Gly	Glu	Pro
20	Val 65	Asn	Ala	Ala	Asp	Ala 70	Ala	Ala	Leu	Glu	His 75	Asp	Lys	Ala	Tyr	Asp 80
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	Asp	Ala	Glu	Phe 100	Gln	Glu	Arg	Leu	Gln 105	Glu	Asp	Thr	Ser	Phe 110	Gly	Gly
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35	Leu 	Gly 130	Leu	Val	Glu	Glu	Gly 135	Ala	Lys	Thr	Ala	Pro 140	Gly	ГЛЗ	Lys	Arg
	Pro 145	Val	Glu	Gln	Ser	Pro 150	Gln	Glu	Pro	qeA	Ser 155	Ser	Ser	Gly	Ile	Gly 160
40	ГÀЗ	Thr	Gly	Gln	Gln 165	Pro	Ala	Lys	Lys	Arg 170	Leu	Asn	Phe	Gly	Gln 175	Thr
45	Gly	Asp	Ser	Glu 180	Ser	Val	Pro	Asp	Pro 185	Gln	Pro	Leu	Gly	Glu 190	Pro	Pro
	Ala	Ala	Pro 195	Ser	Gly	Leu	Gly	Pro 200	Asn	Thr	Met	Ala	Ser 205	Gly	Gly	Gly
50	Ala	Pro 210	Met	Ala	Asp	Asn	Asn 215	Glu	Gly	Ala	qzA	Gly 220	Val	Gly .	Asn	Ser
	Ser 225	Gly	Asn	Trp	His	Cys 230	Asp	Ser	Thr		Leu 235	Gly	Asp	Arg		Ile 240

	Thr	Thr	Ser	Thr	Arg 245	Thr	Trp	Ala	Leu	Pro 250	Thr	Tyr	Asn	Asn	His 255	Leu
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10	Thr	Tyr	Phe 275	Gly	Tyr	Ser	Thr	Pro 280	Trp	Gly	Tyr	Phe	Asp 285	Phe	Asn	Arg
15	Phe	His 290	Суз	His	Phe	Ser	Pro 295	Arg	Asp	Trp	Gln	Arg 300	Leu	Ile	Asn	Asn
	Asn 305	Trp	Gly	Phe	Arg	Pro 310	Lys	Arg	Leu	Asn	Phe 315	Lys	Leu	Phe	Asn	11e 320
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	Pro	туг	Val 355	Leu	Gly	Ser	Ala	His 360	Gln	eſà	Cys	Leu	Pro 365	Pro	Phe	Pro
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45	Asp	Arg	Leu 435	Met	Asn	Pro	Leu	Ile 440	Asp	Gln	Tyr	Leu	Tyr 445	Tyr	Leu	Val
	Arg	Thr 450	Gln	Thr	Thr	Gly	Thr 455	Gly	Gly	Thr	Gln	Thr 460	Leu	Ala	Phe	Ser
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55	Gly	Pro	Cys	Tyr	Arg 485	Gln	Gln	Arg	Val	Ser 490	Thr	Thr	Thr	Asn	Gln 495	Asn

	2	Asn	Asn	Sei	Asr 500	Pho	e Ala	a Tr	p Thi	50	y Ala 5	a Aļ	a Ly:	s Pho	e Ly: 510		u Asn
5	C	€ly	Arg	Asp 515	Ser	Lev	ı Met	L Ası	520	o Gly	y Val	l Ala	a Met	525		r His	s Lys
10	2	rsb	Asp 530	Asp	Asp	Arg	g Phe	9 Phe 535	Pro	Sei	Sei	: Gly	7 Va] 540		ı Ile	Phe	e Gly
45	1 5	ys 45	Gln	Gly	Ala	Gly	7 Asn 550	Asp	Gly	Val	. Asp	туг 555		Glr	val	. Leu	1 Ile 560
15	Т	hr	qeA	Glu	Glu	Glu 565	Ile	Lys	Ala	Thr	570		Val	. Ala	Thr	Glu 575	Glu
20	т	yr	Gly	Ala	Val 580	Ala	Ile	Asn	Asn	Gln 585	Ala	Ala	Asn	Thr	Gln 590		Gln
	T	hr	elà	Leu 595	Val	His	Asn	Gln	Gly 600	Val	Ile	Pro	Gly	Met 605	Val	Trp	Gln
25	A	sn j	Arg 610	Asp	Val	Tyr	Leu	Gln 615	Gly	Pro	Ile	Trp	Ala 620	Lys	Ile	Pro	His
30	T) 6:	hr ) 25	<b>q</b> e/	Gly	Asn	Phe	His 630	Pro	Ser	Pro	Leu	Met 635	Gly	Gly	Phe	Gly	Leu 640
	Ly	ys I	lis	Pro	Pro	Pro 645	Gln	Ile	Leu	Ile	<b>Lys</b> 650	Asn	Thr	Pro	Val	Pro 655	Ala
35	As	I qe	?ro	Pro	Leu 660	Thr	Phe	Asn	Gln	Ala 665	Lys	Leu	Asn	Ser	Phe 670	Ile	Thr
40	G)	n T	'yr	Ser 675	Thr	Gly	Gln	Val	Ser 680	Val	Glu	Ile	Glu	Trp 685	Glu	Leu	Gln
45	Ly	's G	lu 1 90	Asn	Ser	Lys	Arg	Trp 695	Asn	Pro	Glu	Ile	<b>Gln</b> 700	Tyr	Thr	Ser	Asn
45	Ту 70	r T 5	yr 1	ГÀЗ	Ser	Thr	Asn 710	Val	Asp	Phe		Val 715	Asn	Thr	Glu	Gly	Val 720
50	Ту	r S	er (	<b>3</b> lu	Pro :	Arg 725	Pro	Ile	ejà .		Arg 730	Tyr	Leu	Thr		Asn 735	Leu
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<400> 98

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Gly Tyr Lys Tyr Leu Gly Pro Phe Asn Gly Leu Asp Lys Gly Glu  So Val Asn Ala Ala Asp Ala Ala Ala Leu Glu His Asp Lys Ala Tyr  So Gln Gln Leu Lys Ala Gly Asp Asn Pro Tyr Leu Arg Tyr Asn His  So S		Glu	Gly	Ile		Glu	Trp	Trp	Asp		Lys	Pro	G17	Ala		Lys	Pro	
Val Asn Ala Ala Asp Ala Ala Ala Leu Glu His Asp Lys Ala Tyr .  65	10	Lуз	Ala		Gln	Gln	Lys	Gln		Asp	Gly	Arg	Gly		. Val	Leu	Pro	
Gln Gln Leu Lys Ala Gly Asp Asn Pro Tyr Leu Arg Tyr Asn His see See Gly Asp Asn Pro Tyr Leu Arg Tyr Asn His see See Gly Asp Asn Pro Tyr Leu Arg Tyr Asn His see See Gly Asp Asp Asn Pro Tyr Leu Arg Tyr Asn His see See Gly Asp Asp Asp Asp Asp Pro Tyr Leu Arg Tyr Asn His see See Gly Asp Asp Asp Asp Asp Asp Pro Tyr Leu Arg Tyr Asn His see See See See See See See See See Se	15	Gly		Lys	Tyr	Leu	Gly		Phe	Asn	Gly	Leu		Lys	еĵу	Glu	Pro	
Asp Ala Glu Phe Gln Glu Arg Leu Gln Glu Asp Thr Ser Phe Gly Glu Ilo Gly Arg Ala Val Phe Gln Ala Lys Lys Arg Val Leu Glu Ili		Val 65	Asn	Ala	Ala	Asp		Ala	Ala	Leu	Glu		Азр	Lys	Ala	Tyr	Asp 80	
Asn Leu Gly Arg Ala Val Phe Gln Ala Lys Lys Arg Val Leu Glu 115  Leu Gly Leu Val Glu Glu Gly Ala Lys Thr Ala Pro Gly Lys Lys Arg 130  Pro Val Glu Gln Ser Pro Gln Glu Pro Asp Ser Ser Ser Gly Ile Glu 145  Lys Thr Gly Gln Gln Pro Ala Lys Lys Arg Leu Asn Phe Gly Gln 7 175  Gly Asp Ser Glu Ser Val Pro Asp Pro Gln Pro Leu Gly Glu Pro Files  Ala Ala Pro Ser Gly Leu Gly Pro Asn Thr Met Ala Ser Gly Gly Gly Glo 195  Ala Pro Met Ala Asp Asn Asn Glu Gly Ala Asp Gly Val Gly Asn Ser Gly Asn Trp His Cys Asp Ser Thr Trp Leu Gly Asp Arg Val I	20	Gln	Gln	Leu	Lys		Gly	Asp	Asn	Pro		Leu	Arg	Tyr	Asn		Ala	
Leu Gly Leu Val Glu Glu Gly Ala Lys Thr Ala Pro Gly Lys Lys A 130    Pro Val Glu Gln Ser Pro Gln Glu Pro Asp Ser Ser Ser Gly Ile G 145    Lys Thr Gly Gln Gln Pro Ala Lys Lys Arg Leu Asn Phe Gly Gln T 165    Gly Asp Ser Glu Ser Val Pro Asp Pro Gln Pro Leu Gly Glu Pro E 180    Ala Ala Pro Ser Gly Leu Gly Pro Asn Thr Met Ala Ser Gly Gly G 195    Ala Pro Met Ala Asp Asn Asn Glu Gly Ala Asp Gly Val Gly Asn S 210    Ser Gly Asn Trp His Cys Asp Ser Thr Trp Leu Gly Asp Arg Val I	25	Asp	Ala	Glu	Phe 100	Gln	Glu	Arg	Leu		Glu	Asp	Thr	Ser		Gly	GJA	
Pro Val Glu Gln Ser Pro Gln Glu Pro Asp Ser Ser Ser Gly Ile Glass Thr Gly Gln Gln Pro Asp Ser Ser Ser Gly Ile Glass Thr Gly Gln Gln Pro Ala Lys Lys Arg Leu Asn Phe Gly Gln To 175  Gly Asp Ser Glu Ser Val Pro Asp Pro Gln Pro Leu Gly Glu Pro E 180  Ala Ala Pro Ser Gly Leu Gly Pro Asn Thr Met Ala Ser Gly Gly Glass Ser Gly 195  Ala Pro Met Ala Asp Asn Asn Glu Gly Ala Asp Gly Val Gly Asn Ser Gly Asn Trp His Cys Asp Ser Thr Trp Leu Gly Asp Arg Val I		Asn	Leu	Gly 115	Arg	Ala	Val	Phe		Ala	Lys	Lys	Arg		Leu	Glu	Pro	
Lys Thr Gly Gln Gln Pro Ala Lys Lys Arg Leu Asn Phe Gly Gln 7 175  Gly Asp Ser Glu Ser Val Pro Asp Pro Gln Pro Leu Gly Glu Pro F 180  Ala Ala Pro Ser Gly Leu Gly Pro Asn Thr Met Ala Ser Gly Gly Gly 195  Ala Pro Met Ala Asp Asn Asn Glu Gly Ala Asp Gly Val Gly Asn S 215  Ser Gly Asn Trp His Cys Asp Ser Thr Trp Leu Gly Asp Arg Val T	30	Leu	Gly 130	Leu	Val	Glu	Glu	Gly 135	Ala	Lys	Thr	Ala		Gly	Lys	Lys	Arg	
Gly Asp Ser Glu Ser Val Pro Asp Pro Gln Pro Leu Gly Glu Pro E 180  Ala Ala Pro Ser Gly Leu Gly Pro Asn Thr Met Ala Ser Gly	35	Pro 145	Val	Glu	Gln	Ser		Gln	Glu	Pro	Asp		Ser	Ser	Gly	Ile	Gly 160	
Ala Ala Pro Ser Gly Leu Gly Pro Asn Thr Met Ala Ser Gly		Lys	Thr	Gly	Gln		Pro	Ala	Lys	Lys		Leu	Asn	Phe	Gly		Thr	
Ala Pro Met Ala Asp Asn Asn Glu Gly Ala Asp Gly Val Gly Asn S 210 215 220 Ser Gly Asn Trp His Cys Asp Ser Thr Trp Leu Gly Asp Arg Val I	40	Gly	Asp	Ser	Glu 180	Ser	Val	Pro	Asp		Gln	Pro	Leu	Gly		Pro	Pro	
210 215 220  Ser Gly Asn Trp His Cys Asp Ser Thr Trp Leu Gly Asp Arg Val I	45	Ala	Ala	Pro 195	Ser	Gly	Leu			Asn	Thr	Met	Ala		Gly	Gly	ely	
Ser GIV Asn Trp His Cys Asp Ser Thr Trp Leu Gly Asp Arg Val I		Ala	Pro 210	Met	Ala	Asp	Asn		Glu	Gly .	Ala			Val	Gly	Asn	Ser	
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	Tyr	Lys	Gln	11e 260		Asn	Gly	Thr	Ser 265		Gly	Ser	Thr	Asn 270	_	Asn
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	Asn 305		Gly	Phe	Arg	Pro 310	Lys	Arg	Leu	Asn	Phe 315		Leu	Phe	Asn	Ile 320
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	Pro	Tyr	Val 355	Leu	Gly	Ser	Ala	His 360	Gln	Gly	Суз	Leu	Pro 365	Pro	Phe	Pro
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35	Gly 385	Ser	Gln	Ala	Leu	Gly 390	Arg	Ser	Ser	Phe	Tyr 3 <b>9</b> 5	Cys	Leu	Glu	Tyr	Phe 400
	Pro	Ser	Gln	Met	Pro 405	Arg	Thr	GJÀ	Asn	Asn 410	Phe	Gln	Phe	Ser	Tyr 415	Thr
40	Phe	Glu	Asp	Val 420	Pro	Phe	His	ser	Ser 425	Tyr	Ala	His	Ser	Gln 430	Ser	Leu
45	Asp	Arg	Leu 435	Met	Asn	Pro	Leu	Ile 440	Asp	Gln	Tyr	Leu	Tyr 445	Tyr	Leu	Val
	Arg	Thr 450	Gln	Thr	Thr	Gly	Thr 455	Gly	Gly	Thr	Gln	Thr 460	Leu	Ala	Phe	Ser
50	Gln 465	Ala	Gly	Pro	Ser	Ser 470	Met	Ala	Asn	Gln	Ala 475	Arg	Asn	Trp	Val	Pro 480
55	СſА	Pro	Сув	TYr	Arg 485	Gln	Gln	Arg	Val	Ser 490	Thr	Thr	Thr	Asn	Gln 495	Asn

	Asn	Asn	Ser	Asn 500	Phe	Ala	Trp	Thr	Gly 505	Ala	Ala	Lys	Phe	Lys 510		Asn
5	Gly	Arg	Asp 515	Ser	Leu	Met	Asn	Pro 520	Gly	Val	Ala	Met	Ala 525		His	Lys
10	Asp	Asp 530	Asp	Asp	Arg	Phe	Phe 535	Pro	Ser	Ser	GŢĀ	Val 540	Leu	Ile	Phe	Gly
	Lys 545		Gly	Ala	elà	Asn 550	Asp	Gly	Val	Asp	Tyr 555	Ser	Gln	Val	Leu	Ile 560
15	Thr	Asp	Glu	Glu	Glu 565	Ile	Lys	Ala	Thr	Asn 570	Pro	Val	Ala	Thr	Glu 575	<b>Gl</b> u
20	Tyr	Gly	Ala	Val 580	Ala	Ile	Asn	Asn	Gln 585	Ala	Ala	Asn	Thr	Gln 590	Ala	<b>Gl</b> n
95	Thr	Gly	Leu 595	Val	His	Asn	Gln	Gly 600	Val	Ile	Pro	Gly	Met 605	Val	Trp	Gln
25	Asn	Arg 610	Asp	Val	Tyr	Leu	Gln 615	Gly	Pro	Ile	Trp	Ala 620	Lys	Ile	Pro	His
30	Thr 625	Asp	Gly	Asn	Phe	His 630	Pro	Ser	Pro	Leu	Met 635	Gly	Gly	Phe	Gly	Leu 640
05	Lys	His	Pro	Pro	Pro 645	Gln	Ile	Leu	Ile	Lys 650	Asn	Thr	Pro	Val	Pro 655	Ala
35	Asp	Pro	Pro	Leu 660	Thr	Phe	Asn	Gln	Ala 665	Lys	Leu	Asn	Ser	Phe 670	Ile	Thr
40	Gln	Tyr	Ser 675	Thr	Gly	Gln	Val	Ser 680	Val	Glu	Ile	Glu	Trp 685	Glu	Leu	Gln
45	Lys	Glu 690	Asn	Ser	Lys	Arg	Trp 695	Asn	Pro	Glu	Ile	Gln 700	Tyr	Thr	Ser	Asn
40	Туг 705	Tyr	Гуз	Ser	Thr	Asn 710	Val	Asp	Phe	Ala	Val 715	Asn	Thr	Glu	Gly	<b>Val</b> 720
50	Туг	Ser	<b>Gl</b> u		Arg 725	Pro	Ile	Gly		Arg 730	Tyr	Leu	Thr	Arg	Asn 735	Leu
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<213> capsid protein of AAV serotype, clone 43.20

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	Glu	Gly	Ile	Arg 20	Glu	Trp	Trp	qeA	Leu 25	Ľуs	Pro	Gly	Ala	Pro 30	Lys	Pro
10	ГЛЗ	Ala	Asn 35	Gln	Gln	Lys	Gln	Asp 40	Asp	Gly	Arg	Gly	Leu 45	Val	Leu	Pro
15	Gly	Tyr 50	Lys	Tyr	Leu	Gly	Pro 55	Phe	Asn	Gly	Leu	Asp 60	Lys	Gly	Glu	Pro
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25	Aap	Ala	Glu	Phe 100	Gln	Glu	Arg	Leu	Gln 105	Glu	Asp	Thr	Ser	Phe 110	Gly	Gly
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35	Leu 145	Val	Glu	Gln	Ser	Pro 150	Gln	Glu	Pro	Asp	ser 155	Ser	Ser	Gly	Ile	Gly 160
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	Phe	His 290	Cys	His	Phe	Ser	Pro 295	Arg	Asp	Trp	Gln	Arg 300	Leu	Ile	Asn	Asn
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	Asn	Leu	Thr	Ser 340	Thr	Val	Gln	Val	Phe 345	Thr	Asp	Ser	Glu	Tyr 350	Gln	Leu
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	Gly 385	Ser	Gln	Ala	Leu	Gly Gly	Arg	Ser	Ser	Phe	Tyr 395	Суз	Leu	Glu	Туг	Phe 400
35	Pro	Ser	Gln	Met	Leu 405	Arg	Thr	Gly	Asn	Asn 410	Phe	Gln	Phe	Ser	Tyr 415	Thr
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	Asp	Arg	Leu 435					Ile 440							Leu	Val
45	Arg	Thr 450	Gln	Thr	Thr	Gly	Thr 455	Gly	Gly	Thr	Gln	Thr 460	Leu	Ala	Phe	Ser
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	GЈУ	Pro	Cys	Tyr	Arg 485	Gln	Gln	Arg	Val	Ser 490	Thr	Thr	Thr	Asn	Gln 495	Asn

_	Ası	Asn	Ser	Asn 500	Phe	Ala	Trp	Thr	Gly 505	Ala	Ala	Lys	Phe	Lys 510	Leu	Asn
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10	Ası	Asp 530		qeA	Arg	Phe	Phe 535	Pro	ser	Ser	Gly	Val 540	Leu	Ile	Phe	Gly
15	Lys 545	Gln	Gly	Ala	Gly	Asn 550	Asp	Gly	Val	Asp	Tyr 555	Ser	Gln	Val	Leu	Ile 560
	Thr	Asp	Glu	Glu	Glu 565	Ile	Lys	Ala	Thr	Asn 570	Pro	Val	Ala	Thr	Glu 575	Glu
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	Asr	Arg 610	Asp	Val	Tyr	Leu	Gln 615	Gly	Pro	Ile	Trp	Ala 620	ГÀЗ	Ile	Pro	His
30	Thr 625	Asp	Gly	Asn	Phe	His 630	Pro	Ser	Pro	Leu	Met 635	Gly	GŢÀ	Phe	Gly	Leu 640
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	Азр	Pro	Pro	Leu 660	Thr	Phe	Asn	Gln	Ala 665	Lys	Leu	Asn	Ser	Phe 670	Ile	Thr
40	Gln	Tyr	Ser 675	Thr	Gly	Gln	Val	<b>Ser</b> 680	Val	Glu	Ile	Glu	Trp 685	Glu	Leu	Gln
45	Lys	Glu 690	Asn	Ser	Lys	Arg	Trp 695	Asn	Pro	Glu	Ile	Gln 700	Tyr	Thr	Ser	Asn
	Tyr 705	Tyr	Lys	Ser	Thr	Asn 710	Val	Asp	Phe	Ala	Val 715	Asn	Thr	Glu	Gly	Val 720
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5	Glu	Gly	Ile	Arg 20	Glu	Trp	Trp	Asp	Leu 25	Lys	Pro	Gly	Ala	Pro 30	Lys	Pro
10	Lys	Ala	Asn 35	Gln	Gln	Lys	Gln	Asp 40	Asp	Gl <b>y</b>	Arg	еĵу	Leu 45	Val	Leu	Pro
	Gly	Tyr 50	ГЛЗ	Tyr	Leu	Gly	Pro 55	Phe	Asn	elà	Leu	<b>Аз</b> р 60	Lys	Gly	Glu	Pro
15	Val 65	Asn	Ala	Ala	Азр	Ala 70	Ala	Ala	Leu	Glu	His 75	Asp	Lys	Ala	Tyr	Asp 80
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	Asp	Ala	Glu	Phe 100	Gln	Glu	Arg	Leu	Gln 105	Glu	Ąsp	Thr	Ser	Phe 110	Gly	Gly
25	Asn	Leu	Gly 115	Arg	Ala	Val	Phe	Gln 120	Ala	Lys	Lys	Arg	Val 125	Leu	Glu	Pro
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	Pro 145	Val	Glu	Gln	Ser	Pro 150	Gln	Glu	Pro	Ąsp	Ser 155	Ser	Ser	Gly	Ile	Gly 160
35	Lys	Ser	Gly	Gln	Gln 165	Pro	Ala	Lys	Lys	Arg 170	Leu	Asn	Phe	Gly	Gln 175	Thr
40	GŢĀ	Asp	Ser	Glu 180	Ser	Val	Pro	Asp	Pro 185	Gln	Pro	Leu	GГÀ	Glu 190	Pro	Pro
	Glu	Ala	Pro 195	Ser	Gly	Leu	Gly	Pro 200	Asn	Thr	Met	Ala	Ser 205	Gly	Gly	Gly
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#### 40 Claims

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- 1. A method of identifying unknown adeno-associated virus (AAV) sequences in a sample suspected of containing AAV from a latent infection, said method comprising the steps of:
  - (a) subjecting the sample containing DNA to amplification via polymerase chain reaction (PCR) using a first set of primers which specifically amplify a first AAV region comprising at least 250 bp of AAV capsid nucleic acid sequences, said first region having a variable sequence flanked by at least 18 base pairs of highly conserved sequence at its 5' end and at least 18 base pairs of highly conserved sequence at its 3' end, said base pairs being highly conserved relative to an alignment of at least AAV1, AAV2, AAV3, AAV4, AAV5 and AAV6;
  - (b) optionally subjecting the DNA to further amplification using a second set of primers which specifically amplify a second region which comprises the first region of AAV sequences and sequences which are 5' to the first region, such that AAV 5' extension sequences which anneal to the 5' end of the AAV sequences amplified by the primers for the first region are obtained;
  - (c) optionally subjecting the DNA to further amplification using a third set of primers which specifically amplify a third region which comprises the first region of AAV sequences and sequences which are 3' to the first region, such that AAV 3' extension sequences which anneal to the 3' end of the AAV sequences amplified by the primers for the first region are obtained,

each of said second and third regions being predetermined based upon the alignment of the nucleic acid sequences of at least AAV1, AAV2, AAV3. AAV4, AAV5 and AAV6, and each of said regions comprising nucleic acid sequences which are highly conserved over at least 18 base pairs at the 5' end, optionally variable sequences in the middle, and sequences which are highly conserved over at least 18 base pairs at the 3' end of the sequences of the region, relative to the sequences of at least AAV1, AAV2, AAV3, AAV4, AAV5 and AAV6; and each of the sets of primers consisting of a 5' primer and a 3' primer; the presence of amplified sequences indicating the presence of an AAV in the sample, and a comparison of differences between the amplified sequences and the sequences of AAV1, AAV2, AAV3, AAV4, AAV5 and AAV6 indicating the presence of an unknown AAV.

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- 2. A method according to claim 1, wherein the comparison comprises the step of comparing restriction enzyme patterns for the amplified sequences to restriction enzyme patterns of AAV1, AAV2, AAV3, AAV4, AAV5 and AAV6.
- 3. A method according to claim 1 or claim 2, wherein step (a) amplifies the full-length capsid gene.

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4. A method according to any of claims 1 to 3, wherein the amplified sequences comprise the AAV capsid gene and the AAV rep gene.

5. A method according to any of claims 1 to 4, wherein the DNA has been extracted from cells, cell culture, tissue, tissue culture or biological fluids.

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6. A method according to any of claims 1 to 5, wherein the first region is highly conserved over at least about 25 base pairs at the 5' end of the region, the 3' end of the region or both.

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7. A method according to claim 6, wherein the first region is highly conserved over at least about 30 base pairs at the 5' end of the region, the 3' end of the region or both.

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8. A method according to any of claims 1 to 7, wherein the highly conserved sequences of the first region have at least 80% identity among the aligned AAVs at the 5' end of the region, the 3' end of the region or both.

9. A method according to claim 8, wherein the highly conserved sequences of the first region have at least 90% identity among the aligned AAVs at the 5' end of the region, the 3' end of the region or both.

10. A method according to any of claims 1 to 9, wherein the variable sequences in the middle of the first region have less than 70% identity among the aligned AAVs.

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11. A method according to any of claims 1 to 10, wherein the first region spans about bp 2800 to about 3200 of AAV 1, SEQ ID NO:6, and corresponding base pairs in other AAVs.

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12. A method according to claim 11, wherein the first region is 257 bp spanning bp 2886 to about 3143 of AAV 1, SEQ ID NO:6, and corresponding base pairs in other AAVs.

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13. A method according to any of claims 1 to 5, wherein the primers axe AV1ns, having the sequence ofnncleotides 1398 to 1423 of SEQ ID NO:6. and AV2cas, having the sequence of SEQ ID NO:7.

14. A method according to claim 1 or claim 2, wherein the first set of primers allows isolation of full-length adeno-associated virus capsid sequences from a sample, the first set of primers comprising a 5' primer directed to a region located in the middle of an AAV rep gene, based on a predetermined conserved region, and a 3' primer directed to a region downstream of an AAV cap gene, based

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15. A method according to any of claims 1 to 14, wherein the sample comprises AAV integrated into the chromosome.

16. A method according to any of claims 1 to 15, wherein the sample comprises human tissue.

on a predetermined conserved region of AAV.

- 17. A method according to any of claims 1 to 16, wherein the sample contains proviral AAV sequences.
- 18. A method according to any of claims 1 to 17, wherein the first region is a signature region.

- 19. A method according to any of claims 1 to 18, wherein the base pairs of the highly conserved sequences are highly conserved relative to an alignment of AAVs 1,2,3,4,5 and 6 and AAVs isolated from geese and ducks.
- 20. A method according to any of claims 1 to 19, wherein the variable sequence is a hypervariable sequence.

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- 21. A method according to any of claims 1 to 20, wherein the first region comprises up to 10 kilobasepairs in length.
- 22. A method according to claim 21, wherein the first region comprises a 3-1 kilobase pair fragment comprising the full-length cap sequence.
- 23. A kit for detecting the presence of an unknown adeno-associated virus (AAV) in a sample from cellular DNA suspected of containing a latent AAV infection, said kit comprising:
  - (a) a first set of primers which specifically amplify a first region comprising 250 bp of AAV capsid nucleic acid sequences, said first region having at least 18 base pairs of highly conserved sequence at its 5' end, a variable sequence, and at least 18 base pairs of highly conserved sequence at its 3' end, said base pairs being highly conserved relative to an alignment of at least AAV1, AAV2, AAV3, AAV4, AAV5 and AAV6;
  - (b) optionally a second set of primers specific for a second region of the AAV nucleic acid sequences which comprises the first region of AAV sequences and sequences which are 5' to the first region, such that AAV 5' extension sequences which anneal to the 5' end of the AAV sequences amplified by the primers for the first region are obtained;
  - (c) optionally a third set of primers which specifically amplify a third region which comprises the first region of AAV sequences and sequences which are 3' to the first region, such that AAV 3' extension sequences which anneal to the 3' end of the AAV sequences amplified by the primers for the first region are obtained;

each of said second and third regions being predetermined based upon the alignment of the nucleic acid sequences of at least AAV1, AAV2, AAV3, AAV4, AAV5 and AAV6, and each of said regions comprising nucleic acid sequences which are highly conserved over at least 18 base pairs at the 5' end, optionally variable sequences in the middle, and sequences which are highly conserved over at least 18 base pairs at the 3' end of the sequences of the region, relative to the sequences of at least AAV1, AAV2, AAV3, AAV4, AAV5 and AAV6; each of the sets of primers consisting of a 5' primer and a 3' primer, each of said primers comprising at least 15 nucleotides complementary to its respective highly conserved sequence and having exact identity with its respective

- 24. A kit according to claim 23, wherein the 5' primer and/or the 3' primer comprises at least 18 nucleotides.
  - 25. A kit according to claim 24, wherein the 5' primer and/or the 3' primer comprises 25 nucleotides.

highly conserved sequence over at least 5 base pairs in its 3' end.

- **26.** A kit according to any of claims 23 to 25, wherein the 5' primer and/or the 3' primer comprises at least 9 base pairs of exact identity at its 3' end.
  - 27. A kit according to claim 26, wherein the 5' primer and/or the 3' primer comprises at least 18 base pairs of exact identity at its 3' end.
- 28. A kit according to any of claims 23 to 27, wherein the first set of primers allows isolation of full-length adeno-associated virus capsid sequences from a sample, the first set of primers comprising a 5' primer directed to a region located in the middle of an AAV rep gene, based on a predetermined conserved region of AAV, and a 3' primer directed to a region downstream of an AAV cap gene, based on a predetermined conserved region of AAV.
  - 29. A kit according to claim 23, wherein the 5' primer has a sequence comprising GCTGCGTCAACTGGACCAATGA-GAAC, which corresponds to nt 1398 to 1423 of SEQ ID NO:6.
  - **30.** A kit according to claim 23, wherein the 3' primer has a sequence comprising CGCAGAGACCAAAGTTCAACT-GAAACGA, which corresponds to the nucleotides complementary to 4462-4435 of SEQ ID NO:7.
    - 31. A kit according to any of claims 23 to 30, wherein the sample comprises AAV integrated into the chromosome.

#### Patentansprüche

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- Verfahren zur Identifizierung unbekannter Sequenzen von adeno-assoziiertem Virus (AAV) in einer Probe, von der man annimmt, daß sie von einer latenten Infektion herrührendes AAV enthält, wobei man in den folgenden Verfahrenschritten
  - (a) die DNA-haltige Probe einer Amplifikation über eine Polymerasekettenreaktion (PCR) unter Verwendung eines ersten Primersatzes, mit dem spezifisch ein mindestens 250 Bp AAV-Capsid-Nukleinsäuresequenzen umfassender erster AAV-Bereich amplifiziert wird, wobei dieser erste Bereich eine an ihrem 5'-Ende von mindestens 18 Basenpaaren hochkonservierter Sequenz und an ihrem 3'-Ende von mindestens 18 Basenpaaren hochkonservierter Sequenz aufweist, wobei die Basenpaare relativ zu einer vergleichenden Anordnung von mindestens AAV1, AAV2, AAV3, AAV4, AAV5 und AAV6 hochkonserviert sind, aussetzt
  - (b) gegebenenfalls die DNA einer weiteren Amplifikation unter Verwendung eines zweiten Primersatzes, mit dem spezifisch ein zweiter Bereich, der den ersten Bereich von AAV-Sequenzen sowie 5' zum ersten Bereich liegende Sequenzen umfaßt, amplifiziert wird, aussetzt, so daß 5'-AAV-Verlängerungssequenzen, die in einer Annealing-Reaktion an das 5'-Ende der mit den Primern für den ersten Bereich amplifizierten AAV-Sequenzen binden, erhalten werden,
  - (c) gegebenenfalls die DNA einer weiteren Amplifikation unter Verwendung eines dritten Primersatzes, mit dem spezifisch ein dritter Bereich, der den ersten Bereich von AAV-Sequenzen sowie 3' zum ersten Bereich liegende Sequenzen umfaßt, amplifiziert wird, aussetzt, so daß 3'-AAV-Verlängerungssequenzen, die in einer Annealing-Reaktion an das 3'-Ende der mit den Primern für den ersten Bereich amplifizierten AAV-Sequenzen binden, erhalten werden,
- wobei der zweite und der dritte Bereich jeweils auf der Grundlage der vergleichenden Anordnung der Nukleinsäuresequenzen von mindestens AAV1, AAV2, AAV3, AAV4, AAV5 und AAV6 vorbestimmt sind und die Bereiche relativ zu den Sequenzen von mindestens AAV1, AAV2, AAV3, AAV4, AAV5 und AAV6 jeweils am 5'-Ende der Sequenzen des Bereichs über mindestens 18 Basenpaare hochkonservierte Nukleinsäuresequenzen, in der Mitte gegebenenfalls variable Sequenzen und am 3'-Ende über mindestens 18 Basenpaare hochkonservierte Sequenzen umfassen und
  - die Primersätze jeweils aus einem 5'-Primer und einem 3'-Primer bestehen, das Vorhandensein amplifizierter Sequenzen das Vorhandensein eines AAV in der Probe anzeigt, und ein Vergleich der Unterschiede zwischen den amplifizierten Sequenzen und den Sequenzen von AAV1, AAV2, AAV3, AAV4,
  - 2. Verfahren nach Anspruch 1, wobei der Vergleich den Schritt des Vergleichens von Restriktionsenzymmustern für die amplifizierten Sequenzen mit Restriktionsenzymmustern von AAV1, AAV2, AAV3, AAV4, AAV5 und AAV6 umfaßt.
- 40 3. Verfahren nach Anspruch 1 oder 2, wobei in Schritt (a) das Capsid-Gen in voller Länge amplifiziert wird.

AAV5 und AAV6 das Vorhandensein eines unbekannten AAV anzeigt.

- 4. Verfahren nach einem der Ansprüche 1 bis 3, wobei die amplifizierten Sequenzen das AAV-Capsid-Gen und das AAV-rep-Gen umfassen.
- 5. Verfahren nach einem der Ansprüche 1 bis 4, wobei die DNA aus Zellen, Zellkultur, Gewebe, Gewebekultur oder biologischen Flüssigkeiten extrahiert wurde.
  - **6.** Verfahren nach einem der Ansprüche 1 bis 5, wobei der erste Bereich über mindestens etwa 25 Basenpaare am 5'-Ende oder/und am 3'-Ende des Bereichs hochkonserviert ist.
  - 7. Verfahren nach Anspruch 6, wobei der erste Bereich über mindestens etwa 30 Basenpaare am 5'-Ende oder/und am 3'-Ende des Bereichs hochkonserviert ist.
- 8. Verfahren nach einem der Ansprüche 1 bis 7, wobei die hochkonservierten Sequenzen des ersten Bereichs unter den vergleichend angeordneten AAVs eine Identität von mindestens 80% am 5'-Ende oder/und am 3'-Ende des Bereichs aufweisen.
  - 9. Verfahren nach Anspruch 8, wobei die hochkonservierten Sequenzen des ersten Bereichs unter den vergleichend

angeordneten AAVs eine Identität von mindestens 90% am 5'-Ende oder/und am 3'-Ende des Bereichs aufweisen.

- 10. Verfahren nach einem der Ansprüche 1 bis 9, wobei die variablen Sequenzen in der Mitte des ersten Bereichs unter den vergleichend angeordneten AAVs eine Identität von weniger als 70% aufweisen.
- 11. Verfahren nach einem der Ansprüche 1 bis 10, wobei der erste Bereich von etwa Bp 2800 bis etwa 3200 von AAV1, SEQ ID NO:6, und den entsprechenden Basenpaaren in anderen AAV reicht.
- **12.** Verfahren nach Anspruch 11, wobei es sich bei dem ersten Bereich um 257 Bp handelt, die von Bp 2886 bis etwa 3143 von AAV1, SEQ ID NO:6, und den entsprechenden Basenpaaren in anderen AAV reichen.

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- 13. Verfahren nach einem der Ansprüche 1 bis 5, wobei es sich bei den Primern um AV1ns mit der Sequenz der Nukleotide 1398 bis 1423 der SEQ ID NO:6 sowie um AV2cas mit der Sequenz der SEQ ID NO:7 handelt.
- 14. Verfahren nach Anspruch 1 oder Anspruch 2, wobei der erste Primersatz die Isolierung von Capsidsequenzen in voller L\u00e4nge von adeno-assoziiertem Virus aus einer Probe gestattet, wobei der erste Primersatz einen auf einen in der Mitte eines AAV-rep-Gens liegenden Bereich auf der Grundlage eines vorbestimmten konservierten Bereichs gerichteten 5'-Primer sowie einen auf einen stromabw\u00e4rts von einem AAV-cap-Gen liegenden Bereich auf der Grundlage eines vorbestimmten konservierten Bereichs von AAV gerichteten 3'-Primer umfa\u00e4t.
  - 15. Verfahren nach einem der Ansprüche 1 bis 14, wobei die Probe in das Chromosom integriertes AAV umfaßt.
  - 16. Verfahren nach einem der Ansprüche 1 bis 15, wobei die Probe menschliches Gewebe umfaßt.
  - 17. Verfahren nach einem der Ansprüche 1 bis 16, wobei die Probe provirale AAV-Sequenzen enthält.
  - **18.** Verfahren nach einem der Ansprüche 1 bis 17, wobei es sich bei dem ersten Bereich um einen Signaturbereich handelt.
  - 19. Verfahren nach einem der Ansprüche 1 bis 18, wobei die Basenpaare der hochkonservierten Sequenzen relativ zu einer vergleichenden Anordnung von AAV 1, 2, 3, 4, 5 und 6 und aus Gans und Ente isolierten AAV hochkonserviert sind.
- 20. Verfahren nach einem der Ansprüche 1 bis 19, wobei es sich bei der variablen Sequenz um eine hypervariable Sequenz handelt.
  - 21. Verfahren nach einem der Ansprüche 1 bis 20, wobei der erste Bereich eine Länge von bis zu 10 Kilobasenpaaren umfaßt.
  - **22.** Verfahren nach Anspruch 21, wobei der erste Bereich ein die cap-Sequenz in voller Länge umfassendes Fragment von 3,1 Kilobasenpaaren umfaßt.
- 23. Kit zum Nachweis des Vorhandenseins eines unbekannten adeno-assoziierten Virus (AAV) in einer Probe aus zellulärer DNA, von der man annimmt, daß sie eine latente AAV-Infektion enthält, wobei der Kit umfaßt:
  - (a) einen ersten Primersatz, mit dem spezifisch ein 250 Bp AAV-Capsid-Nukleinsäuresequenzen umfassender erster AAV-Bereich amplifiziert wird, wobei dieser erste Bereich an seinem. 5'-Ende mindestens 18 Basenpaare hochkonservierter Sequenz, eine variable Sequenz und an seinem 3'-Ende mindestens 18 Basenpaare hochkonservierter Sequenz aufweist, wobei die Basenpaare relativ zu einer vergleichenden Anordnung von mindestens AAV1, AAV2, AAV3, AAV4, AAV5 und AAV6 hochkonserviert sind,
  - (b) gegebenenfalls einen für einen zweiten Bereich der AAV-Nukleinsäuresequenzen, der den ersten Bereich von AAV-Sequenzen sowie 5' zum ersten Bereich liegende Sequenzen umfaßt, spezifischen zweiten Primersatz, so daß 5'-AAV-Verlängerungssequenzen, die in einer Annealing-Reaktion an das 5'-Ende der mit den Primern für den ersten Bereich amplifizierten AAV-Sequenzen binden, erhalten werden,
  - (c) gegebenenfalls einen dritten Primersatz, mit dem spezifisch ein dritter Bereich, der den ersten Bereich von AAV-Sequenzen sowie 3' zum ersten Bereich liegende Sequenzen umfaßt, amplifiziert wird, so daß 3'-AAV-Verlängerungssequenzen, die in einer Annealing-Reaktion an das 3'-Ende der mit den Primern für den ersten

Bereich amplifizierten AAV-Sequenzen binden, erhalten werden,

wobei der zweite und der dritte Bereich jeweils auf der Grundlage der vergleichenden Anordnung der Nukleinsäuresequenzen von mindestens AAV1, AAV2, AAV3, AAV4, AAV5 und AAV6 vorbestimmt sind und die Bereiche relativ zu den Sequenzen von mindestens AAV1, AAV2, AAV3, AAV4, AAV5 und AAV6 jeweils am 5'-Ende der Sequenzen des Bereichs über mindestens 18 Basenpaare hochkonservierte Nukleinsäuresequenzen, in der Mitte gegebenenfalls variable Sequenzen und am 3'-Ende über mindestens 18 Basenpaare hochkonservierte Sequenzen umfassen,

die Primersätze jeweils aus einem 5'-Primer und einem 3'-Primer bestehen, wobei jeder Primer mindestens 15 zur hochkonservierten Sequenz des jeweils anderen Primers komplementäre Nukleotide umfaßt und an seinem 3'-Ende über mindestens 5 Basenpaare eine genaue Identität mit der hochkonservierten Sequenz des jeweils anderen Primers aufweist.

- 24. Kit nach Anspruch 23, wobei der 5'-Primer und/oder der 3'-Primer mindestens 18 Nukleotide umfaßt.
- 25. Kit nach Anspruch 24, wobei der 5'-Primer und/oder der 3'-Primer mindestens 25 Nukleotide umfaßt.
- **26.** Kit nach einem der Ansprüche 23 bis 25, wobei der 5'-Primer und/oder der 3'-Primer an seinem 3'-Ende mindestens 9 Basenpaare genauer Identität umfaßt.
- 27. Kit nach Anspruch 26, wobei der 5'-Primer und/oder der 3'-Primer an seinem 3'-Ende mindestens 18 Basenpaare genauer Identität umfaßt.
- 28. Kit nach einem der Ansprüche 23 bis 27, wobei der erste Primersatz die Isolierung von Capsidsequenzen in voller Länge von adeno-assoziiertem Virus aus einer Probe gestattet, wobei der erste Primersatz einen auf einen in der Mitte eines AAV-rep-Gens liegenden Bereich auf der Grundlage eines vorbestimmten konservierten Bereichs von AAV gerichteten 5'-Primer sowie einen auf einen stromabwärts von einem AAV-cap-Gen liegenden Bereich auf der Grundlage eines vorbestimmten konservierten Bereichs von AAV gerichteten 5'-Primer umfaßt.
  - 29. Kit nach Anspruch 23, wobei der 5'-Primer eine GCTGCGTCAACTGGACCAATGAGAAC umfassende Sequenz aufweist, die Nt 1398 bis 1423 der SEQ ID NO:6 entspricht.
  - **30.** Kit nach Anspruch 23, wobei der 3'-Primer eine CGCAGAGACCAAAGTTCAACTGAAACGA umfassende Sequenz aufweist, die den zu 4462-4435 der SEQ ID NO:7 komplementären Nukleotiden entspricht.
  - 31. Kit nach einem der Ansprüche 23 bis 30, wobei die Probe in das Chromosom integriertes AAV umfaßt.

#### 40 Revendications

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- 1. Procédé pour identifier des séquences de virus associés à l'adénovirus (VAA) inconnus dans un échantillon dont on suspecte qu'il contient des VAA provenant d'une infection latente, ledit procédé comprenant les étapes :
  - (a) de soumission de l'échantillon contenant l'ADN à une amplification via une réaction de polymérase en chaîne (PCR) en utilisant une première série d'amorces qui amplifient spécifiquement une première région de VAA comprenant au moins 250 pb des séquences d'acides nucléiques de capside de VAA, ladite première région présentant une séquence variable adjacente à au moins 18 paires de bases d'une séquence hautement conservée en son extrémité 5' et à au moins 18 paires de bases d'une séquence hautement conservée en son extrémité 3', lesdites paires de bases étant hautement conservées par rapport à un alignement d'au moins VAA1, VAA2, VAA3, VAA4, VAA5 et VAA6;
  - (b) éventuellement de soumission de l'ADN à une autre amplification en utilisant une deuxième série d'amorces qui amplifient spécifiquement une deuxième région qui comprend la première région de séquences des VAA et des séquences qui sont côté 5' par rapport à la première région, de telle manière qu'on obtient des séquences d'extension 5' de VAA qui hybrident sur l'extrémité 5' des séquences de VAA amplifiées par les amorces pour la première région ;
  - (c) éventuellement de soumission de l'ADN à une autre amplification utilisant une troisième série d'amorces qui amplifient spécifiquement une troisième région qui comprend la première région de séquences de VAA et

les séquences qui sont situées côté 3' par rapport à la première région, de telle manière qu'on obtient des séquences d'extension 3' de VAA qui hybrident sur l'extrémité 3' des séquences de VAA amplifiées par les amorces pour la première région,

5 chacune desdites deuxième et troisième régions étant prédéterminée sur base de l'alignement des séquences d'acides nucléiques d'au moins VAA1, VAA2, VAA3, VAA4, VAA5 et VAA6, et chacune desdites régions comprenant des séquences d'acides nucléiques qui sont hautement conservées sur au moins 18 paires de bases en l'extrémité 5', des séquences éventuellement variables au centre et des séquences qui sont hautement conservées sur au moins 18 paires de bases en l'extrémité 3' des séquences de la région, par rapport aux séquences d'au moins 10 VAA1, VAA2, VAA3, VAA4, VAA5 et VAA6; et

chacune des séries d'amorces étant constituée par une amorce 5' et une amorce 3'; la présence de séquences amplifiées indiquant la présence d'un VAA dans l'échantillon et une comparaison des différences entre les séquences amplifiées et les séquences des VAA1, VAA2, VAA3, VAA4, VAA5 et VAA6 indiquant la présence d'un VAA inconnu.

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- 2. Procédé selon la revendication 1, dans lequel la comparaison comprend l'étape de comparaison de modèles d'enzymes de restriction pour les séquences amplifiées à des modèles d'enzymes de restriction des VAA1, VAA2, VAA3, VAA4, VAA5 et VAA6.
- 20 3. Procédé selon la revendication 1 ou 2, dans lequel l'étape (a) amplifie toute 1a longueur du gène cap.
  - 4. Procédé selon l'une quelconque des revendications 1 à 3, dans lequel les séquences amplifiées comprennent le gène cap du VAA et le gène rep du VAA.
- 25 5. Procédé selon l'une quelconque des revendications 1 à 4, dans lequel l'ADN a été extrait de cellules, d'une culture cellulaire, de tissu, d'une culture de tissu ou de fluides biologiques.
  - 6. Procédé selon l'une quelconque des revendications 1 à 5, dans lequel la première région est hautement conservée sur au moins 25 paires de base en l'extrémité 5' de la région, en l'extrémité 3' de la région ou les deux.

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7. Procédé selon la revendication 6, dans lequel la première région est hautement conservée sur au moins 30 paires de base en l'extrémité 5' de la région, en l'extrémité 3' de la région ou les deux.

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autres VAA.

8. Procédé selon l'une quelconque des revendications 1 à 7, dans lequel les séguences hautement conservées de la première région présentent une identité d'au moins 80% avec les VAA alignés en l'extrémité 5' de la région, l'extrémité 3' de la région ou les deux.

9. Procédé selon la revendication 8, dans lequel les séquences hautement conservées de la première région présentent une identité d'au moins 90% avec les VAA alignés en l'extrémité 5' de la région, l'extrémité 3' de la région ou les deux.

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10. Procédé selon l'une quelconque des revendications 1 à 9, dans lequel les séguences variables au centre de la première région présentent une identité inférieure à 70% avec les VAA alignés.

11. Procédé selon l'une quelconque des revendications 1 à 10, dans lequel la première région s'étend de la paire de

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bases 2800 à environ 3200 du VAA 1, SEQ ID NO:6, et les paires de bases correspondantes dans les autres VAA. 12. Procédé selon la revendication 11, dans lequel la première région représente 257 paires de bases, s'étendant de

la paire de bases 2886 à environ 3143 du VAA1, SEQ ID NO:6, et les paires de bases correspondantes dans les

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13. Procédé selon l'une quelconque des revendications 1 à 5, dans lequel les amorces sont des AV1ns, présentant la séquence des nucléotides 1398 à 1423 de la SEQ ID NO:6, et des AV2cas, présentant la séquence de la SEQ ID NO:7.

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14. Procédé selon la revendication 1 ou 2, dans lequel la première série d'amorces permet l'isolement de toute la longueur de séquences de capside du virus associé à l'adénovirus d'un échantillon, la première série d'amorces comprenant une amorce 5' dirigée sur une région localisée au centre d'un gène rep du VAA, sur base d'une région prédéterminée conservée et une amorce 3', dirigée sur une région en aval d'un gène cap du VAA, basée sur une

région prédéterminée conservée du VAA.

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- 15. Procédé selon l'une quelconque des revendications 1 à 14, dans lequel l'échantillon comprend un VAA intégré dans le chromosome.
- 16. Procédé selon l'une quelconque des revendications 1 à 15, dans lequel l'échantillon comprend du tissu humain.
- 17. Procédé selon l'une quelconque des revendications 1 à 16, dans lequel l'échantillon contient des séquences de VAA provirales.
- 18. Procédé selon l'une quelconque des revendications 1 à 17, dans lequel la première région est une région de signature.
- 19. Procédé selon l'une quelconque des revendications 1 à 18, dans lequel les paires de bases des séquences hautement conservées sont hautement conservées par rapport à un alignement des VAA 1,2,3,4,5 et 6 et des VAA isolés à partir d'oies et de canards.
- **20.** Procédé selon l'une quelconque des revendications 1 à 19, dans lequel la séquence variable est une séquence hypervariable.
- 21. Procédé selon l'une quelconque des revendications 1 à 20, dans lequel la première région comprend jusqu'à 10 kilopaires de bases en longueur.
  - 22. Procédé selon la revendication 21, dans lequel la première région comprend un fragment de 3,1 kilopaires de bases comprenant toute la longueur de la séquence du capside.
  - 23. Kit pour détecter la présence d'un virus associé à l'adénovirus (VAA) inconnu dans un échantillon d'ADN cellulaire dont on suspecte qu'il contient une infection latente par un VAA, ledit kit comprenant:
    - (a) une première série d'amorces qui amplifient spécifiquement une première région comprenant 250 paires de bases de séquences d'acides nucléiques d'un capside de VAA, ladite première région présentant au moins 18 paires de bases d'une séquence hautement conservée en son extrémité 5', une séquence variable et au moins 18 paires de base d'une séquence hautement conservée en son extrémité 3', lesdites paires de bases étant hautement conservées par rapport à un alignement d'au moins VAA1, VAA2, VAA3, VAA4, VAA5 et VAA6; (b) éventuellement une deuxième série d'amorces spécifiques d'une deuxième région des séquences d'acides nucléiques de VAA qui comprend la première région des séquences de VAA et des séquences qui se situent côté 5' par rapport à la première région, de manière à obtenir des séquences d'extension 5' des VAA qui hybrident sur l'extrémité 5' des séquences de VAA amplifiées par les amorces pour la première région, qui com-
    - (c) eventuellement une troisieme serie d'amorces qui amplifient specifiquement une troisieme region, qui comprend la première région de séquences de VAA et des séquences qui se situent côté 3' par rapport à la première région, de manière à obtenir des séquences d'extension 3' de VAA qui hybrident sur l'extrémité 3' des séquences de VAA amplifiées par les amorces de la première région;

chacune desdites deuxième et troisième région étant prédéterminée sur base de l'alignement des séquences d'acides nucléiques d'au moins les VAA1, VAA2, VAA3, VAA4, VAA5 et VAA6, et chacune desdites régions comprenant des séquences d'acides nucléiques qui sont hautement conservées sur au moins 18 paires de bases en l'extrémité 5', éventuellement des séquences variables au centre et des séquences qui sont hautement conservées sur au moins 18 paires de bases en l'extrémité 3' des séquences de la région, par rapport aux séquences au moins des VAA1, VAA2, VAA3, VAA4, VAA5 et VAA6;

- chacune des séries d'amorces étant constituée par une amorce 5' et une amorce 3', chacune desdites amorces comprenant au moins 15 nucléotides complémentaires à sa séquence respective hautement conservée et présentant une identité exacte avec sa séquence respective hautement conservée sur au moins 5 paires de bases en son extrémité 3'.
- 24. Kit selon la revendication 23, dans lequel l'amorce 5' et/ou l'amorce 3' comprend au moins 18 nucléotides.
- 25. Kit selon la revendication 24, dans lequel l'amorce 5' et/ou l'amorce 3' comprend 25 nucléotides.
- 26. Kit selon l'une quelconque des revendications 23 à 25, dans lequel l'amorce 5' et/ou l'amorce 3' comprend au moins

9 paires de bases d'identité exacte en son extrémité 3'.

- 27. Kit selon la revendication 26, dans lequel l'amorce 5' et/ou l'amorce 3' comprend au moins 18 paires de bases d'identité exacte en son extrémité 3'.
- 28. Kit selon l'une quelconque des revendications 23 à 27, dans lequel la première série d'amorces permet l'isolement de toute la longueur des séquences de capside d'un virus associé à l'adénovirus d'un échantillon, la première série d'amorces comprenant une amorce 5' dirigée sur une région localisée au centre d'un gène rep d'un VAA, basée sur une région prédéterminée conservée d'un VAA et une amorce 3' dirigée sur une région en aval d'un gène cap d'un VAA, basée sur une région prédéterminée conservée d'un VAA.
- 29. Kit selon 1a revendication 23, dans lequel l'amorce 5' présente une séquence comprenant GCTGCGTCAACTG-GACCAATGAGAAC, ce qui correspond aux nucléotides 1398 à 1423 de la SEQ ID NO:6.
- **30.** Kit selon la revendication 23, dans lequel l'amorce 3' présente une séquence comprenant CGCAGAGACCAAAGTT-CAACTGAAACGA, qui correspond aux nucléotides complémentaires à 4462-4435 de la SEQ ID NO:7.
  - **31.** Kit selon l'une quelconque des revendications 23 à 30, dans lequel l'échantillon comprend un VAA intégré dans le chromosome.